

Pezeshkian, Zahra, Nobili, Stefania, Peyravian, Noshad, Shojaee, Bahador, Nazari, Haniye, Soleimani, Hiva, Asadzadeh-Aghdaei, Hamid, Ashrafian Bonab, Maziar, Nazemalhosseini-Mojarad, Ehsan and Mini, Enrico (2021) Insights into the Role of Matrix Metalloproteinases in Precancerous Conditions and in Colorectal Cancer. Cancers. ISSN 2072-6694

Downloaded from: http://sure.sunderland.ac.uk/id/eprint/14260/

Usage guidelines

sure@sunderland.ac.uk.





Remiero

Insights into the Role of Matrix Metalloproteinases in Precancerous Conditions and in Colorectal Cancer

Zahra Pezeshkian¹, Stefania Nobili^{2,3}, Noshad Peyravian¹, Bahador Shojaee¹, Haniye Nazari⁴, Hiva Soleimani⁵, Hamid Asadzadeh-Aghdaei¹, Maziar Ashrafian Bonab⁶, Ehsan Nazemalhosseini-Mojarad^{7,*} and Enrico Mini^{8,9,*}

- Basic and Molecular Epidemiology of Gastrointestinal Disorders Research Center, Research Institute for Gastroenterology and Liver Diseases, Shahid Beheshti University of Medical Sciences, Tehran 19835-178, Iran; zahrapezeshkian@yahoo.com (Z.P.); peyravian.n@iums.ac.ir (N.P.); bahadorshojaee@ufl.edu (B.S.); hamid.asadzadeh@sbmu.ac.ir (H.A.-A.)
- Department of Neurosciences, Imaging and Clinical Sciences, "G. D'Annunzio" University of Chieti-Pescara, 66100 Chieti, Italy; stefania.nobili@unich.it
- 3 Center for Advanced Studies and Technology (CAST), University "G. d'Annunzio" Chieti-Pescara, 66100 Chieti, Italy
- Department of Microbiology, Faculty of Advanced Science and Technology, Tehran Medical Science, Islamic Azad University, Tehran 19395/1495, Iran; hani7311926@gmail.com
- ⁵ Department of General Biology, Faculty of Fundamental Science, Islamic Azad University of Shahr-E-Qods, Tehran 37515- 374, Iran; M_hivi@yahoo.com
- ⁶ School of Medicine, University of Sunderland, City Campus, Chester Road, Sunderland SR1 3SD, UK; maziar.bonab@sunderland.ac.uk
- Gastroenterology and Liver Diseases Research Center, Research Institute for Gastroenterology and Liver Diseases, Shahid Beheshti University of Medical Sciences, Tehran 19835-178, Iran
- 8 Department of Health Sciences, University of Florence, 50139 Florence, Italy
- 9 DENOTHE Excellence Center, University of Florence, 50139 Florence, Italy
- * Correspondence: E.nazemalhosseini@sbmu.ac.ir (E.N.-M.); enrico.mini@unifi.it (E.M.)

Simple Summary: Colorectal cancer (CRC) is one of the most common cancer worldwide. CRC is derived from polyps and many factors, such as Matrix Metalloproteinases (MMPs) can gain the progression of colorectal carcinogenesis. Many investigations have indicated the role of MMPs in CRC development while there is not enough knowledge about the function of MMPs in precancerous conditions. This review summarizes the current information about the role of MMPs in polyps and CRC progression.

Abstract: Colorectal cancer (CRC) is the third and second cancer for incidence and mortality worldwide, respectively, and is becoming prevalent in developing countries. Most CRCs derive from polyps, especially adenomatous polyps, which can gradually transform into CRC. The family of Matrix Metalloproteinases (MMPs) plays a critical role in the initiation and progression of CRC. Prominent MMPs, including MMP-1, MMP-2, MMP-7, MMP-8, MMP-9, MMP-12, MMP-13, MMP-14, and MMP-21, have been detected in CRC patients, and the expression of most of them correlates with a poor prognosis. Moreover, many studies have explored the inhibition of MMPs and targeted therapy for CRC, but there is not enough information about the role of MMPs in polyp malignancy. In this review, we discuss the role of MMPs in colorectal cancer and its pathogenesis

Keywords: Matrix Metalloproteinases (MMPs); polyp; colorectal cancer; TIMPs; MMP polymorphisms; MMP targeting

Citation: Pezeshkian, Z.; Nobili, S.; Peyravian, N.; Shojaee, B.; Nazari, H.; Soleimani, H.; Asadzadeh-Aghdaei, H.; Bonab, M.A.; Nazemalhoseini-Mojarad, E.; Mini, E. Insights into the Role of Matrix Metalloproteinases in Precancerous Conditions and in Colorectal Cancer. *Cancers* 2021, 13, 6226. https://doi.org/10.3390/cancers1 3246226

Academic Editor: Heike Allgayer

Received: 14 November 2021 Accepted: 7 December 2021 Published: 10 December 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).

Cancers **2021**, 13, 6226 2 of 21

1. Introduction

At approximately 11% of all diagnosed cancer cases, CRC is the third most common cancer and the second most lethal cancer worldwide [1,2]. It is today well known that several factors contribute to the CRC pathogenesis, driving complex genetic and epigenetic processes that, ultimately, transform normal colonic mucosa to cancerous tissue [3]. CRC may initiate from benign polyps with the mucosal origin and can develop into carcinoma. Colorectal polyps, especially adenomas, are proliferative lesions that have been defined as the precursor of CRC. Therefore, the early detection and removal of these polyps can interrupt the progression of the adenoma-carcinoma sequence [4,5].

Many molecular signaling pathways are involved in CRC initiation and progression, such as ERK/MAPK, TGF- β , PI3K/Akt, Src/FAK, and β -catenin pathways. These pathways can promote the hallmarks of cancer such as inflammation, angiogenesis, metastasis, and invasion, also via the activation and overexpression of MMPs [6,7]. Thus, MMPs have been suggested as potential prognostic factors for the malignancy risk of colorectal polyps. MMPs are proteolytic enzymes implicated in the degradation of stromal connective tissues and of the extracellular matrix (ECM), a complex network that plays a key role in sustaining signaling transduction and thus cancer development and progression [8]. As such, MMPs have key roles in tumor initiation, progression, and metastasis and can affect tumor cell behavior by cleaving proapoptotic agents and producing an aggressive phenotype [9]. Because of these roles, MMPs have been detected as biomarkers in CRC progression [10]. A new challenge in CRC treatment is finding an effective pharmacological and therapeutic method for suppression of MMPs and targeted therapy of CRC [11]. This review will deal with the role of MMPs in colorectal carcinogenesis from colorectal polyps to CRC.

2. CRC Pathogenesis and Molecular Classification

Colorectal polyps result from atypical cell proliferation in the colorectal tissue. Based on histological and morphological features, colorectal polyps are divided into neoplastic (adenoma) and non-neoplastic (hyperplastic, hamartomatous, and inflammatory) types [5,12]. Neoplastic polyps, also known as adenomatous polyps, are subclassified by their histological characteristics as tubular, villous, or tubulovillous adenomas. Previous investigations demonstrated that approximately 5–10% of neoplastic polyps are villous adenomas and most of them show dysplasia. Approximately 10–15% of neoplastic polyps show morphological features of both villous and tubular types [13]. Adenomas are not usually transformed to carcinoma, but there is evidence that the adenoma-carcinoma sequence originates from adenomatous polyps [14]. Also, hyperplastic polyps may possess malignancy potential [15]. CRC is caused by the misregulation of some oncogenes such as *KRAS* and *c-MYC* and tumor suppressor genes such as *P53* and APC, which control cellular signal transduction [16–18].

2.1. Molecular Mechanism of CRC

Specific features characterize CRC and its pathogenesis based on genetic, epigenetic, and transcriptomic factors. Three main molecular abnormalities are involved in CRC carcinogenesis:

- A. Microsatellite instability (MSI): it consists of mutations in DNA mismatch repair (MMR) genes such as *MSH2*, *MLH1*, *PMS2*, *MLH3*, *MSH3*, *PMSI*, and *EXO1*; MSI is rare in polyps but it is always found in serrated polyps and about 15–20% of all CRC cases are derived from MSI [19,20].
- B. Chromosomal instability (CIN): this abnormality is identified in 85% of CRC cases and consists of a gain (1q, 7p, 8q, 13q, 2pq) or loss (8q, 15q, 17p, 18p) of chromosomal genes, activation of proto-oncogenes (*KRAS*, *SRC*, *c-MYC*), and inactivation of tumor suppressor genes (*P53*, *APC*) [21].

Cancers 2021, 13, 6226 3 of 21

C. CpG Islands Methylator Phenotype (CIMP): these regions, located in the gene promoter, could disturb the activation of tumor suppressor genes. CIMP phenotype is represented by hypermethylation of CpG dinucleotides and premalignant serrated polyps are correlated with CIMP [22,23]

2.2. Molecular Classification Based on Transcriptomic Analysis

Based on gene expression profiles, CRC has been classified into subgroups with distinct molecular and clinical features [24].

- A. Consensus molecular subtype (CMS) classification: CMS classification provides biological insight into metastatic colorectal cancer (mCRC) carcinogenesis and predicts CRC prognosis [25].
 - o CMS1 (14%) indicates MSI, CIMP, and BRAF mutation and immune activation.
 - CMS2 (37%) shows Wingless-Type MMTR integration site family member (WNT), MYC signaling activation, and epithelial involvement.
 - CMS3 (13%) demonstrates MSI, CIMP, and KRAS mutations and metabolic involvement.
 - CMS4 (23%) includes invasion, metastatic situations, and TGF-β signaling co-activation and angiogenesis. Also, epithelial-mesenchymal transition (EMT) is a crucial event in colorectal carcinogenesis and is involved in CMS4 status. EMT can result in advanced-stage CRC, poor patient survival, and worst clinical features [26,27] and CMS4 subgroup shows the most unfavorable prognosis.
- B. CRC intrinsic subtypes (CRIS): CRIS is a unique classification exclusively based on the cancer cell-specific transcriptome of CRC since the extrinsic factors of the stroma have not been analyzed. It classifies CRC into five novel transcriptional groups that, thus, further clarify biological understanding of CRC heterogeneity.
 - o CRIS-A is enriched for *BRAF*-mutated MSI tumors and *KRAS*-mutated MSS tumors that are without targeted therapeutic options.
 - CRIS-B is related to invasive tumors with poor prognosis and high TGF-ß signaling. CRIS-B is unconnected to the CMS4 mesenchymal subtype, which also indicates aggressive tumors with TGF-ß pathway activation.
 - CRIS-C is dependent on EGFR signals and is sensitive to anti-EGFR monoclonal antibody treatment.
 - CRIS-D shows IGF2 overexpression. This occurrence has been involved in desensitization to the EGFR blockade in patients with *KRAS* wild-type tumors.
 - CRIS-E indicates *KRAS*-mutated, Paneth cell-like CIN tumors refractory to anti-EGFR antibody treatment [28].

3. Structure and Function of MMPs

MMPs are a family of zinc-dependent endopeptidases consisting of a propeptide sequence, a catalytic domain, a hinge region, and a hemopexin (PEX) domain [29]. The propeptide domain is highly conserved and can regulate the sequence that interacts with Zn²⁺. Also, cystine within this area permits the MMPs to be in the active or inactive status [30]. The catalytic domain possesses a conserved zinc-binding motif which, in the active condition, will disconnect from the propeptide domain. Movement between the catalytic and PEX domain is done via hinge regions [29]. According to their structural domains, MMPs have been categorized into collagenase, gelatinase, stromelysin, matrilysin, and membrane-bound MMPs (MT-MMPs) [31,32].

MMPs play a crucial role in the remodeling of the ECM by digestion of ECM components, stimulation of cell surface proteins. Also, they can control the activity of other proteinases, growth factors, chemokines, and cell receptors, and moderate many biological functions [33]. MMPs can regulate cellular growth, migration, survival, and adhesion in biological and pathological statuses (Table 1, Figure 1). Due to the MMP's key roles, the dysregulation of their expression levels and their activation lead cancerous cells to

Cancers 2021, 13, 6226 4 of 21

proliferation, angiogenesis, survival, invasion, malignant transitions, and immune dysregulation [34–36]. Also, the tissue inhibitors of metalloproteinase (TIMPs) control the activation of MMPs and have a critical action in precancerous conditions, CRC progression, and metastasis (Table 2, Figure 2) [11,37].

MMP Gene	Chromosomal Loca- tion	Enzyme	Substrate
MMP-1	11q22.2	Collagenase-1	Col I,II, III, VII, VIII, X, Gelatin
MMP-8	11q22.2	Collagenase-2	Col I,II, III, VII, VIII, X, Gelatin, Aggrecan
MMP-13	11q22.2	Collagenase-3	Col I,II, III, VII, VIII, X, Gelatin
MMP-2	16q12.2	Gelatinase A	Gelatin, Col I,II, III, IV, VII
MMP-9	20q13.12	Gelatinase B	Gelatin, Col IV, V
MMP-3	11q22.3	Stromelysin-1	Col II, III, IV, IX, X, proteoglycans, fibronectin, laminin, and elastin.
MMP-10	11q22.2	Stromelysin-2	Col II, III, IV, IX, X, proteoglycans, fibronectin, laminin, and elastin
MMP-7	11q22.2	Marilysin-1	Fibronectin, Laminin, Col I, Gelatin
MMP-14	14q11.2	MT-MMP	Gelatin, Fibronectin, Laminin
MMP-12	11q22.2	Metalloelastase	Gelatin, Fibronectin, Col IV
MMP-21	10q26.2	XMMP	Aggrecan

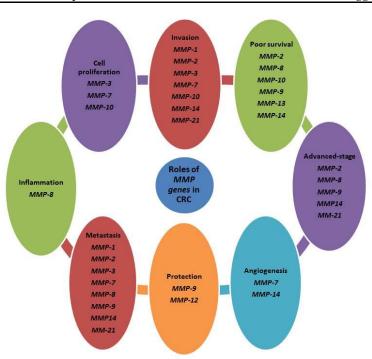


Figure 1. Summary of the prominent *MMP* genes in CRC. MMPs play different functions in CRC.

Cancers **2021**, 13, 6226 5 of 21

Table 2. Summary of Investigations about the Roles of MMP Genes and Proteins in Colorectal Polyps and Cancer.

	Gene/Prote	<u> </u>		
References		Samples	Methods	Results
Huang X, et al., 2021 [38]	sion MMP-7, MMP-9, MMP-11, TIMP-1, TIMP-2, CEA	Human polyps and tumor	Enzyme-linked immuno- sorbent assay	A combined detection model, including MMP-7, TIMP-1, and CEA improved both the specificity and sensitivity for detecting CRC.
Zhou X, et al., 2021 [39]	MMP-7, MMP-9, MMP-11, TIMP-1, TIMP-2, CEA	Human CRC	ELISA and electrochemilumi- nescence immunoassay	The miR 135a was down-regulated and MMP 13 was increased in samples. Combined detection of the two had a good diagnostic effect on the occurrence of CRC.
Rasool M, et al. 2021 [40]	TGF, VEGF, TNF, ILs, MMP-2, 9, 11, and 19	Human polyps and tumor	ELISA	Significant upregulation of MMP-2, MMP-9, MMP-11, and MMP-19 was reported in polyp and colon cancer samples compared with their MMP profile in normal samples.
Barabás L, et al., 2020 [41]	MMP-2, MMP-7, MMP-9, TIMP-1 and TIMP-2	Human ad- enomas, and CRC	ELISA	The serum antigen concentrations of MMP-7, MMP-9, TIMP-1, and TIMP-2 were significantly increased in patients with CRC and adenomas compared with the controls. They were also activated in premalignant adenomas.
Hsieh SL, et al., 2019 [42]	Study of the mecha- nism of carnosine, TIMP-1, and MMP-9	Human HCT-116	MTT assay and qPCR.	The carnosine inhibits the migration and intravasation of human CRC cells. The regulatory mechanism may occur by suppressing NF-κB activity and modulating MMPs and EMT-related gene expression in HCT-116 cells treated with carnosine. MMP-9 mRNA and protein levels were decreased. TIMP-1 mRNA and protein levels were increased.
Kıyak R, et al., 2018 [43]	MMP-7, COX-2, TIMP-1, and CEA protein	Human polyps	ELISA and chemiluminescent enzyme immunometric assay (CEIA)	The plasma TIMP-1 levels were significantly elevated in cancer compared with the

Cancers **2021**, 13, 6226 6 of 21

Fezeshkian ZI, and 14 MMP-7, and MMP-1, and the lowest levels of MMP-1 and MMP-14, and the lowest levels of MMP-1, 7, and 14 compared with the normal group. The MMP-7 mRNA expression was significantly higher in villous adenoma with ligh-grade dysplasia compared with the control group. Association between pathologic stage and immune-reactive score (IRS) polyps and CRC tissue Wernicke AK, et al., 2016 [46] MMP-3 mad MMP-13 has been idenomas and dysplasia compared with the control group. The MMP-13 has been idenomas and limmune-reactive score (IRS) immune-reactive score (IRS) polyps, and CRC tissue Patients' blood, adenoma, by-perplastic polyps, and CRC tissue Patients' stool samples, adenomas, hy-perplastic polyps, and CRC tissue Patients' stool samples, adenomas, hy-perplastic polyps, and CRC tissue Patients' stool samples, adenomas, hy-perplastic polyps, and CRC tissue Patients' stool samples, adenomas, hy-perplastic polyps, and CRC tissue Patients' stool samples, adenomas, hy-perplastic polyps, and CRC tissue Patients' stool samples, adenomas, hy-perplastic polyps, and CRC tissue Patients' stool samples, adenomas with singler than in non-advanced adenomas. Patients' stool samples, adenomas, hy-perplastic polyps, and CRC tissue Patients' stool samples, adenomas with manural tissue levels of MMP-9 may be a new noninvasive marker in CRC.					
Eiró N, et al., 2017 [44] Pezeshkian Z, and 14 Pezeshkian Z, at al., 2017 [45] Pezeshkian Z, and et al., 2017 [45] Pezeshkian Z, and at al., 2016 [46] Wernicke Wernicke AK, et al., 2016 [46] Wernicke Z, and Z, and Z, and Z, and Z, 2016 [47] Pezeshkian Z, and Z,					
Fezeshkian Z, and yerg-A 2017 [44] Pezeshkian Z, and yerg-A 2017 [45] Pezeshkian Z, and yerg-A 2017 [45] Pezeshkian Z, and werd-A, and yerg-A 2017 [45] Pezeshkian Z, and werd-A, and werd-Boliophys Real-time PCR and West-ern-blot, and hyperplastic polyps Real-time PCR and West-ern-blot, and hyperplastic polyps had high levels of both MMP-74, and MMP-74, multiple-villous adenomas had high levels of MMP-17, and 1/4 compared with the normal group. The MMP-7 mRNA expression was significantly higher in villous adenoma with high-grade dysplasia commas including villous, tubular, pared with the control group MMP-7 and VEGF-A are prognostic biomarkers for colorectal adenoma polyp progression to malignancy. Human adenomas and tubulo-villous types, and 20 paired tissue samples. Immunohistochemistry and hyperplastic polyps and cRC. The MMP-13 has been deletified as an excellent marker of high-grade intraepithelial neoplasia and CRC. The strength of the association between pathologic stage and immune-reactive MMP-13 scoring emphasizes its potential for diagnosis in precancerous colorectal lesions. There was a significant correlation between pathologic stage and immune-reactive MMP-13 scoring emphasizes its potential for diagnosis in precancerous colorectal lesions. There was a significant orrelation between palsan and day perplastic polyps, and CRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and CRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and CRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and CRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and cRC tissue perplastic polyps and cRC tissue perplastic polyps, and cRC tissue perplastic polyps, and cRC tissue perplastic polyps, and the power perplast					
Human adenomas and hyperplastic polyps Pezeshkian Z, and 14 2017 [44] Association between Wernicke AK, et al., 2016 [46] Ammahad and MMP-13 and MMP-13 and MMP-13 and MMP-13 and tubulo-villous sdenomas with high-grade dysplasia commas and hyperplastic polyps Association between grade of dysplasia and MMP-13 across the following samples of adenomas and hyperplastic polyps and CRC tissue Patients' Stool samples, Annaha'zi A, et al., 2016 [47] Annaha'zi A, et al., 2016 [48] Annaha'zi Ann					
Eliró N, et al., 2017 [44] and 14 enomas and hyperplastic polyps Pezeshkian Z, et al., 2017 [45] et al., 2016 [46] am MMP-13 arcia A, et al., 2016 [47] am MMP-9, 2016 [47] Amnaha'zi A, et al., 2016 [48] expression Amnaha'zi A, et al., 2016 [48] expression Amnaha'zi A, et al., 2016 [48] expression AmmP-9 and MMP-9, and denomas and hyperplastic polyps, and creating polyps Human adel hyperplastic polyps Real-time PCR and West ern-blot, and a flux lubulo-villous adenomas had high levels of MMP-1, 7, and tubulo-villous adenomas had high levels of MMP-1, 7, and MMP-13, and tubulo-villous yes amples of adenomas including villous, tubulary and and tubulo-villous yes, and and tubulo-villous yes, and significantly higher in villous adenoma with high-grade dysplasia compared with the control group progression to malignancy. The MMP-7 and VECF-A are prognostic biomarkers for colorectal adenoma polyp progression to malignancy. The MMP-13 has been identified as an excellent marker of high-grade intraepithelial neoplasia and CRC. The strength of the association between pathologic stage and immune-reactive MMP-13 scoring emphasizes its potential for diagnosis in precancerous colorectal lesions. Luminex XMAP technology, glatin zymography, western blot, and SNP analysis in 150 blood and tissue evels of MMP-9 levels in pathologic and single than in non-advanced adenomas. Stool MMP-9 was significant Corpared with all the other groups. Stool MMP-9 was significant Corpared with all the other groups. Stool MMP-9 may be a new prophysic marker in CRC.					
Eiró N, et al., 2017 [44] 2017 [44] NAMP-1, 2, 7, 9, 11, 13 and 14 Pezeshkian Z, et al., 2017 [45] Wernicke AK, et al., 2016 [46] Wernicke AK, et al., 2016 [47] Wernicka A, et al., 2016 [47] Annaha'zi A, et al., 2016 [48] Annaha'zi A, et al., 2016 [48] Annaha'zi A, et al., 2016 [48] Pezeshkian Z, et al., 2016 [48] Wernicke Annaha'zi A, et al., 2016 [48] Pezeshkian Z, et al., 2016 [48] Wernicke Annaha'zi A, et al., 2016 [48] Wernicke A, et al., 2016 [48] Wernicke Association between grade of and tubulo-villous types, and enomas and high levels of both MMP-1, and the menbal hyperplastic polyps, and chomas and high levels of both MMP-1, and the menbal hyperplastic polyps and byperplastic polyps, and chomas and high levels of both MMP-1, and the menbal hyperplastic polyps and byperplastic polyps and command high levels of both MMP-1, and thubulo-villous adenomas had high levels of both MMP-1, and the menbal high ern-blot, and byperplastic polyps and enomas and high levels of both MMP-1, and the liquid the normal futulo-villous and menbal polyps and bissue samples. Real-time PCR in Real-time PCR in Stol biopsy samples of adenomaticulus vibular, pared with the control group. Real-time PCR in Real-time PCR in Stol biopsy samples of adenomaticulus vibular, pared with the control group. Human adenomas and alt tubulo-villous types, and significantly higher in villous adenoma with high-grade dysplasia comman high perglastic polyps and tubulo-villous types, and tubulo-villous denoma with high-grade dysplasia comman high perglasia comman high pergl					
Eiró N, et al., 2017 [44] 7, 9, 11, 13 and 14 polyps recommendate encommendate encommendation between grade of AK, et al., 2016 [46] MMP-13 expression Gimeno-Gar cía A, et al., 2016 [47] MMP-9 2016 [48] M			Human ad-		1 11
Pezeshkian Z, and 14 MMP-7 and et al., 2017 [45] Wernicke Wernicke AK, et al., 2016 [46] Wernicke AK, et al., 2016 [47] Wernicke AR, et al., 2016 [48] Wernicke AR, et al., 2016 [47] Wernicke AR, et al., 2016 [48] Association between and tubulo-villous adenoma with biolopsy samples of adeno- mas including villous, tubular, pared with the control group and tubulo-villous adenoma with biolopsy samples of adeno- mas including villous, tubular, pared with the control group and tubulo-villous adenoma with high-grade dysplasia com- mas including villous, tubular, pared with the control group and tubulo-villous adenoma with high-grade dysplasia com- mas including villous, tubular, pared with the control group and tubulo-villous adenoma with high-grade dysplasia com- mas including villous, tubular, pared with the control group and tubulo-villous adenoma with high-grade dysplasia com- mas including villous, tubular, pared with the control group and tubulo-villous adenoma with high-grade dysplasia com- mas including villous, tubular, pared with the control group and tubulo-villous adenoma vibring-14 and vibring-14 and vibring-14 and vibring-14 and vibring-18 and vibring-1	Eiró N. et al		enomas and hyperplastic	Real-time PCR and West-	
Pezeshkian Z, et al., 2017 [45] Wernicke AK, et al., 2016 [46] Gimeno-Gar cia A, et al., 2016 [47] Fezeshkian Real-time PCR in 50 biopsy samples of adenomas including villous, tubular, pared with the control group and tubulo-villous types, and 20 paired tissue samples. Real-time PCR in 50 biopsy samples of adenomas significantly higher in villous adenoma with high-grade dysplasia commas including villous, tubular, pared with the control group and tubulo-villous types, and 20 paired tissue samples. Human adenomas Gimeno-Gar cia A, et al., 2016 [47] Gimeno-Gar cia A, et al., 2016 [47] Fatients' blood, adenomas, hyperplastic polyps, and CRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and CRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and crack at al., 2016 [48] Patients' stool samples, adenomas, hyperplastic polyps, and crack at al., 2016 [48] Patients' stool samples, adenomas, hyperplastic polyps, and crack at al., 2016 [48] Patients' stool samples, adenomas, hyperplastic polyps, and crack at al., 2016 [48] Patients' stool samples, adenomas, hyperplastic polyps, and crack at al., 2016 [48] Patients' stool samples, adenomas, hyperplastic polyps, and crack at al., 2016 [48] Patients' stool samples, adenomas, hyperplastic polyps, and crack at al., 2016 [48] Patients' stool samples, adenomas, hyperplastic polyps, and crack at al., 2016 [48] Patients' stool samples, adenomas high levels of MMP-3 mRNA expression was significantly higher in villous adenoma with high-grade dysplasia commas invision was significantly higher in villous adenoma with high-grade dysplasia commas high levels of MMP-1 and VEGF-A are prognostic biomarkers for colorectal adenoma polyp prognession to malignancy. The MMP-13 habeen identified as an excellent marker in class and immune-reactive score (IRS) The MMP-13 high revis of MMP-13 and typer		7, 9, 11, 13			
Pezeshkian Z, and et al., 2017 [45] Wernicke Wernicke AK, et al., 2016 [46] MMP-13 expression Gimeno-Gar cia A, et al., 2016 [47] Gimeno-Gar Annaha'zi A	. ,	and 14		•	
Pezeshkian Z, and wEGF-A 2017 [45] Association between Wernicke AK, et al., 2016 [46] Egimeno-Gar cia A, et al., 2016 [47] Annaha'zi Annaha'zi Annaha'zi A, et al., 2016 [48] Patients' stool samples, and CRC tissue Patients' stool samples, and tubulo-villous, tubular, pared with the control group and tubulo-villous, tubular, pared with the control group and tubulo-villous, tubular, pared with the control group and tubulo-villous types, and biobios, tubular, pared with the control group and tubulo-villous types, and prognostic biomarkers for colorectal adenoma with high-grade dysplasia commas including villous, tubular, pared with the control group prognostic biomarkers for colorectal adenoma with high-grade dysplasia commas including villous, tubular, pared with the control group prognostic biomarkers for colorectal adenoma prognostic biomarkers for colorectal denoma prognostic biomarkers for colore					_
Pezeshkian Z, et al., 2017 [45] Association between Wernicke AK, et al., 2016 [46] Gimeno-Gar cía A, et al., 2016 [47] Annaha'zi A, et al., 2016 [47] Annaha'zi A, et al., 2016 [48] Pezeshkian Z, et al., 2016 [48] Pezeshkian MMP-9 and Peres Real-time PCR in 50 biopsy samples of adenomas with bigh-grade dysplasia commas including villous, tubular, pared with the control group and tubulo-villous types, and 20 paired tissue samples. Putman adenomas 20 paired tissue samples. Putman adenomas and hyperplastic polyps and 20 paired tissue samples. Immunohistochemistry and hyperplastic polyps and 20 paired tissue samples. Immunohistochemistry and hyperplastic polyps and creation between plasma and tissue levels of MMP-13 scoring emphasizes its potential for diagnosis in precancerous colorectal lesions. There was a significant correlation between plasma and tissue levels of MMP-9. Patients' blood and tissue gelatin zymography, western blot, and SNP analysis in 150 polyps, and CRC tissue Patients' stool samples, adenoma 20 lyp progression to malignancy. The MMP-13 has been identified as an excellent marker of high-grade intraepithelial neoplasia and CRC. The strength of the association between plasma and tissue levels of MMP-9. Patients' blood and tissue gelatin zymography, western blot, and SNP analysis in 150 polyps, and creation between plasma and tissue levels of MMP-9 in CRC was higher than in nealthy controls. Also, MMP-9 in CRC was higher than in nealthy controls. Also, MMP-9 was significantly in it in villous adenoma with high-grade dysplasia componence with the control group in villous adenoma plesson with					-
Pezeshkian Z, and et al., 2017 [45] Wernicke Wernicke AK, et al., 2016 [46] Gimeno-Gar cía A, et al., 2016 [47] Annaha'zi A, et al., 2016 [47] Annaha'zi A, et al., 2016 [48] Patients' blood, adenomas, ples, and corrected A, et al., 2016 [48] Annaha'zi A, et al., 2016 [48] Patients' stool samples of adenomas ples, and tubulo-villous types, and cenomas and hyperplastic polyps, and CRC tissue Real-time PCR in 50 biopsy samples of adenomas including villous, tubular, pared with the control group mas including villous, tubular, pared with the control group progression to malignancy. The MMP-1 and vEGF-A are prognestic biomarkers for colorectal adenoma polyp progression to malignancy. The MMP-13 has been identified as an excellent marker of high-grade intrapily may be a new significantly higher in villous adenoma with higher in villous adenoma villous tubular, pared with the control group into the villous tubular, pared with the control group into the villous tubular, pared with the control group into the villous tubular, pared with the control group into the villous tubular, pared with the control group into the villous tubular, pared with the control group into the villous tubular, pared with the control group into the villous tubular, pared with the control group into the villous tubular, pared with the control group into the villous tubular, pared with the control group into the villous addenoma polyp progression to malignancy. The MMP-13 has been identified as an excellent marker of high-grade intrapilous tubular, pared with the control group into the villous and scale of the minum into the villous and scale					
Pezeshkian Z, and et al., 2017 [45] Wernicke Wernicke AK, et al., 2016 [46] MmP-13 expression Gimeno-Gar cia A, et al., 2016 [47] Annaha'zi Annaha'zi Annaha'zi Annaha'zi Annaha'zi Annaha'zi A, et al., 2016 [48] Patients' stool samples, and cet al., 2016 [48] Patients' stool samples, and cet al., 2016 [48] Patients' stool samples, and cet al., 2016 [48] Real-time PCR in 50 biopsy samples of adenomas with high-grade dysplasia commas including villous, tubular, parade with the control group and tubulo-villous types, and 20 paired tissue samples. Real-time PCR in 50 biopsy samples of adenomas with high-grade dysplasia commas including villous, tubular, parade with the control group and tubulo-villous types, and 20 paired tissue samples. Human adenomas 20 paired tissue samples. Human adenomas and tubulo-villous types, and 20 paired tissue samples. Human adenomas 20 paired tissue samples. Human adenomas and tubulo-villous, tubular, pared with the control group and tubulo-villous types, and and tubulo-villous types, and and tubulo-villous types, and tubulo-villous types, and tubulo-villous types, and and tubulo-villous types, and tubulo-villous types, and mlmp-9 and verification and excellent marker of high-grade dysplasia om and tubulo-villous types, and mlmp-9 and verification and excellent marker of high-grade intraceptive the fifed as an excellent marker of high-grade intraceptive fifed as an excellent marker of high-grade intraceptive fifed as an excellent marker of high-grade intraceptive fifed as an excellent marker of high-grade dysplasia commanders for colorectal alenomas and tits succession to malignancy. The MMP-13 as an excellent marker of high-grade					÷
Pezeshkian Z, and et al., 2017 [45] Werricke Wernicke AK, et al., 2016 [46] Bimeno-Gar cía A, et al., 2016 [47] Annaha'zi Annaha'zi Annaha'zi A, et al., 2016 [48] Patients' stool samples of adenomas including villous, tubular, pared with the control group. and tubulo-villous types, and 20 paired tissue samples. So biopsy samples of adenomas including villous, tubular, pared with the control group. MMP-7 and VEGF-A are prognostic biomarkers for colorectal adenoma polyp progression to malignancy. The MMP-13 has been identified as an excellent marker of high-grade intracepithelial neoplasia and CRC. The strength of the association between pathologic stage and immune-reactive MMP-13 scoring emphasizes its potential for diagnosis in precancerous colorectal lesions. The MMP-13 has been identified as an excellent marker of high-grade intracepithelial neoplasia and CRC. The strength of the association between pathologic stage and immune-reactive MMP-13 scoring emphasizes its potential for diagnosis in precancerous colorectal lesions. Luminex XMAP technology, gelatin zymography, western blot, and SNP analysis in 150 blood and tissue CRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and CRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and CRC tissue Patients' stool samples of adenomas polype prograed with the control groups. The MMP-13 has been identified as an excellent marker of high-grade intracepithelial neoplasia and CRC. The strength of the association between pathologic stage and immune-reactive MMP-13 scoring emphasizes its potential for diagnosis in precancerous colorectal lesions. There was a significant correlation between plasma and tissue levels of MMP-9. Plasma MMP-9 levels in patients with neoplastic lesions were significantly higher than in healthy controls. Also, MMP-9 in CRC compared with all the other groups. Stool MMP-9 may be a new noninvasive marker in CRC				Post Care DCD in	
et al., 2017 [45] et al., VEGF-A and VEGF-A and veger-A and veger-A and veger-A and veger-A and veger-A and veger-A and tubulo-villous types, and 20 paired tissue samples. Association between grade of dysplasia and man and typerplastic polyps expression Association between grade of dysplasia and mMMP-13 expression Arroia A, et al., 2016 [47] Annaha'zi A, et al., 2016 [48] Annaha'zi A, et al., 2016 [48] Patients' stool samples, and 20 paired tissue samples. Immunohistochemistry and hyperplastic polyps	Pezeshkian	1414D 7			
er al., 2017 [45] VEGF-A Association between grade of dysplasia and MP-13 expression Gimeno-Gar cia A, et al., 2016 [47] Annaha'zi A, et al., 2016 [48] Patients' blood, adenomas, hyperplastic polyps, and CRC tissue Patients' stool samples. Patients' stool samples. Patients' blood and tissue Patients' stool samples. Patients' stool samples. Patients' blood and tissue CRC tissue Patients' stool samples. Patients' blood and tissue Patients' blood and tissue CRC tissue Patients' blood and tissue Patients' blood and tissue CRC tissue Patients' blood and tissue CRC tissue Patients' stool samples. Patients' sto	Z,	_	Human ad-		
2017 [45] 20 paired tissue samples. 30 paired tissue samples. 30 paired tissue samples. 4 penbars promayes intitied interpithelial neoplasia description between pairon	et al.,		enomas	_	
Colorectal adenoma polyp progression to malignancy. The MMP-13 has been identified as an excellent marker of high-grade intraepithelial neoplasia and CRC. The strength of the association between grade of dysplasia and polyps polyps and hyperplastic polyps and polyps. Gimeno-Gar cía A, et al., 2016 [47] Gimeno-Gar cía A, et al., 2016 [47] Fatients' blood, adenomas, hyperplastic polyps, and CRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and CRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and CRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and Stool MMP-9 was significantly increased in CRC compared with all the other groups. Stool MMP-9 may be a new ponninyasive marker in CRC	2017 [45]	VLGI -A		* *	
Association Wernicke AK, et al., 2016 [46] Gimeno-Gar cía A, et al., 2016 [47] WMP-9 Climeno-Gar cía A, et al., 2016 [47] Association between dysplasia and MMP-9 Patients' blood, adenomas, hyperplastic polyps, and CRC tissue Patients' blood, adenomas, hyperplastic polyps, and CRC tissue Patients' blood and tissue blood and tissue CRC tissue Patients' blood and tissue cRC tissue Stool MMP-9 was significantly higher than in non-advanced adenomas. Stool MMP-9 was significantly increased in CRC compared with all the other groups. Stool MMP-9 may be a new poninyasive marker in CRC				20 paired tissue samples.	
Association between Wernicke AK, et al., 2016 [46] Gimeno-Gar cía A, et al., 2016 [47] WMP-9 Annaha'zi A, et al., 2016 [48] Association between Human adenomas and hyperplastic polyps Human adenomas and hyperplastic polyps Fatients' blood, adenomas, hyperplastic polyps, and CRC tissue Patients' blood and tissue Patients' blood and tissue CRC tissue Stool MMP-9 was significantly increased in CRC compared with all the other groups. Stool MMP-9 may be a new noninvasive marker in CRC					
Wernicke AK, et al., 2016 [46] Gimeno-Gar cía A, et al., 2016 [47] Amnaha'zi A, et al., 2016 [48] Association between grade of dysplasia and MMP-9 and enomas and hyperplastic polyps Ak, et al., 2016 [47] Bettients' blood, adenomas, hyperplastic polyps, and CRC tissue Patients' blood and tissue CRC tissue Stool MMP-9 was significantly increased in CRC compared with all the other groups. Stool MMP-9 may be a new noninvasive marker in CRC compared with all the other groups. Stool MMP-9 may be a new noninvasive marker in CRC					
Mernicke AK, et al., 2016 [46] Gimeno-Gar cía A, et al., 2016 [47] Annaha'zi A, et al., 2016 [48] Annaha'zi A, et al., 2016 [48] Annaha'zi A, et al., 2016 [48] Association Human adenomas and hyperplastic polyps Human adenomas and hyperplastic polyps Patients' blood, adenomas, hyperplastic polyps, and CRC tissue Patients' blood and tissue Patients' blood and tissue Patients' blood and tissue CRC tissue Patients' blood and tissue Patients' blood and tissue CRC tissue Patients' blood and tissue Stool MMP-9 was significantly higher than in non-advanced adenomas. Patients' stool samples, adenomas, hyperplastic polyps, and CRC tissue Patients' blood and tissue Annaha'zi A, et al., 2016 [48] Patients' stool samples, adenomas, hyperplastic polyps, and CRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and cRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and cRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and cRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and cRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and cRC tissue Patients' blood and tissue Stool MMP-9 was significantly increased in CRC compared with all the other groups. Stool MMP-9 may be a new noninvasive marker in CRC companied with all the other groups.					
Wernicke AK, et al., 2016 [46] where and dysplasia and MMP-13 expression Gimeno-Gar cía A, et al., 2016 [47] where Annaha'zi A, et al., 2016 [48] where Annaha'zi A, et al., 2016 [48] where ARA an Annaha'zi A, et al., 2016 [48] where ARA an Amaha'zi A, et al., 2016 [48] where ARA an		Association	ı.		
Wernicke AK, et al., 2016 [46] and MMP-13 expression Gimeno-Gar cía A, et al., 2016 [47] MMP-9 Annaha'zi A, et al., 2016 [48] Annaha'zi Annaha'zi Annaha'zi Annaha'zi Annaha'zi Annaha'zi Annaha'zi Annaha'zi A, et al., 2016 [48] Annaha'zi Annaha'		_			
AK, et al., 2016 [46] and MMP-13 expression enomas and hyperplastic polyps expression ex	Wernicke				-
2016 [46] and MMP-13 polyps stage and immune-reactive MMP-13 scoring emphasizes its potential for diagnosis in precancerous colorectal lesions. Patients' blood, adenomas, hyperplastic polyps, and CRC tissue Patients' stool samples, adenomas. Patients' stool samples, adenomas, hyperplastic polyps, and CRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and CRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and CRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and CRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and CRC tissue Patients' stool samples, adenomas. Patients' stool samples, adenomas, hyperplastic polyps, and SNP analysis in 150 son, MMP-9 was significantly higher than in healthy controls. Alson MMP-9 was significantly increased in CRC compared with all the other groups. Stool MMP-9 may be a new noninvasive marker in CRC compared with all the other noninvasive marker in CRC c	AK, et al.,	dysplasia and MMP-13	hyperplastic polyps		=
MMP-13 expression MMP-13 scoring emphasizes its potential for diagnosis in precancerous colorectal lesions. There was a significant correlation between plasma and tissue levels of MMP-9. Patients' blood, adenomas, hyperplastic polyps, and CRC tissue Patients' blood, adenomas, hyperplastic polyps, and CRC tissue Patients' stool samples, adenomas. Patients' stool samples, adenomas, hyperplastic polyps, and cRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and ples, adenomas, hyperplastic polyps, and created adenomas. Patients' stool samples, adenomas, hyperplastic polyps, and created adenomas. Patients' stool samples, adenomas, hyperplastic polyps, and created adenomas. Stool MMP-9 was significantly higher than in non-advanced adenomas. Stool MMP-9 was significantly increased in CRC compared with all the other groups. Stool MMP-9 may be a new noninvasive marker in CRC compared with all the other groups.					
Gimeno-Gar cía A, et al., 2016 [47] Annaha'zi A, et al., 2016 [48] Patients' blood, adenomas, hyperplastic polyps, and 2016 [48] Patients' stool samples, adenomas, hyperplastic polyps, and 2016 [48] Patients' stool samples, adenomas, hyperplastic polyps, and 2016 [48] Patients' stool samples, adenomas, hyperplastic polyps, and 2016 [48] Patients' stool samples, adenomas, hyperplastic polyps, and 2016 [48] Patients' stool samples, adenomas, hyperplastic polyps, and 2016 [48]					MMP-13 scoring emphasizes
Gimeno-Gar cía A, et al., 2016 [47] Annaha'zi A, et al., 2016 [48] Annaha'zi A, et al., 2016 [48] Barbara A Patients' blood, adenomas, hyperplastic polyps, and 2016 [48] Barbara B Patients' blood, adenomas, hyperplastic polyps, and 2016 [48] Barbara B Patients' blood, adenomas XMAP technology, plasma MMP-9 levels in patients with neoplastic lesions were significantly higher than in healthy controls. Also, MMP-9 in CRC was higher than in non-advanced adenomas. Barbara B Patients' stool samples, adenomas, hyperplastic polyps, and perplastic polyps, and significant torrelation between plasma and tissue levels of MMP-9. Plasma MMP-9 levels in patients with neoplastic lesions were significantly higher than in healthy controls. Also, MMP-9 in CRC was higher than in non-advanced adenomas. Barbara B Patients' stool samples, adenomas, hyperplastic polyps, and solve perplastic polyps, and solve perplastic polyps, and polyps, and solve perplastic polyps, and solve perplastic polyps, and polyps, and provided the plasma MMP-9 in CRC was higher than in non-advanced adenomas. Barbara B Patients' stool AMP-9 was significant torrelation between plasma and tissue levels of MMP-9 was ignificant torrelation between plasma and tissue levels of MMP-9 was ignificantly higher than in healthy controls. Also, MMP-9 was significantly higher than in non-advanced adenomas. Barbara B Patients' stool AMP-9 was significantly higher than in non-advanced adenomas. Barbara B Patients' stool AMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy cont					
Gimeno-Gar cía A, et al., 2016 [47] Annaha'zi A, et al., 2016 [48] Patients' blood, adenomas, hyperplastic polyps, and CRC tissue Patients' blood, adenomas, hyperplastic polyps, and CRC tissue Patients' blood, adenomas, hyperplastic polyps, and CRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and CRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and compared with all the other groups. Stool MMP-9 may be a new noninvasive marker in CRC relation between plasma and tissue levels of MMP-9. Plasma MMP-9 levels in patients with neoplastic lesions were significantly higher than in healthy controls. Also, MMP-9 in CRC was higher than in non-advanced adenomas. Stool MMP-9 was significant correlation between plasma and tissue levels of MMP-9 levels in patients with neoplastic lesions were significantly higher than in healthy controls. Also, MMP-9 in CRC was higher than in non-advanced adenomas. Stool MMP-9 was significant correlation between plasma and tissue levels of MMP-9 levels in patients with neoplastic lesions were significantly higher than in healthy controls. Also, MMP-9 in CRC was higher than in non-advanced adenomas. Stool MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in					precancerous colorectal le-
Gimeno-Gar cía A, et al., 2016 [47] MMP-9 Annaha'zi A, et al., 2016 [48] Patients' blood, adenomas, hyperplastic polyps, and Annaha'zi A, et al., 2016 [48] Belian Simple Stool samples, and polyps, and plood and tissue stool samples, adenomas, hyperplastic polyps, and stool samples, adenomas, hyperplastic polyps, and plood and tissue stool samples, adenomas, hyperplastic polyps, and stool samples and tissue levels of MMP-9 levels in patients with neoplastic lesions were significantly higher than in healthy controls. Also, MMP-9 in CRC was higher than in non-advanced adenomas. Stool MMP-9 was significantly higher than in non-advanced adenomas. Stool MMP-9 was significantly higher than in non-advanced adenomas. Stool MMP-9 was significantly higher than in non-advanced adenomas. Stool MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in non-advanced adenomas. Stool MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in non-advanced adenomas. Stool MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in					
Gimeno-Gar cía A, et al., 2016 [47] Annaha'zi A, et al., A, et al., A, et al., 2016 [48] Patients' blood, adenomas, hyperplastic polyps, and CRC tissue Patients' blood, adenomas, hyperplastic polyps, and blood and tissue than in healthy controls. Also, MMP-9 in CRC was higher than in non-advanced adenomas. Patients' stool samples, adenomas, hyperplastic polyps, and ples, adenomas, hyperplastic polyps, and ples, adenomas, hyperplastic polyps, and preplastic polyps, and prepl		MMP-9	blood, ade- nomas, hy- perplastic polyps, and	gelatin zymography, western blot, and SNP analysis in 150	Ö
Gimeno-Gar cía A, et al., 2016 [47] MMP-9 perplastic polyps, and CRC tissue Patients' stool samples, adenomas, hyperplastic A, et al., A, et al., 2016 [48] MMP-9 perplastic polyps, and CRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and pless, adenomas, hyperplastic polyps, and CRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and pless, adenomas, hyperplastic polyps, and proplyps, and proply					_
nomas, hyperplastic polyps, and CRC tissue Patients' stool samples, adenomas, hyperplastic A, et al., A, et al., 2016 [48] Annaha'zi A, et al., 2016 [48] Annaha'zi A, et al., 2016 [48] NMMP-9 annomas, hyperplastic polyps, and CRC tissue NMMP-9 blood and tissue than in healthy controls. Also, MMP-9 in CRC was higher than in non-advanced adenomas. Stool MMP-9 was significantly higher than in healthy controls. Also, MMP-9 in CRC was higher than in non-advanced adenomas. Stool MMP-9 was significantly higher than in healthy controls. Also, MMP-9 in CRC was higher than in non-advanced adenomas. Stool MMP-9 was significantly higher than in healthy controls. Also, MMP-9 in CRC was higher than in non-advanced adenomas. Stool MMP-9 was significantly higher than in healthy controls. Also, MMP-9 in CRC was higher than in non-advanced adenomas. Stool MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in healthy contr					
cía A, et al., 2016 [47] MMP-9 perplastic polyps, and CRC tissue Patients' stool samples, adenomas, hyperplastic polyps, and Annaha'zi A, et al., 2016 [48] MMP-9 Patients' stool samples, adenomas, hyperplastic polyps, and polyps, western tients with neoplastic lesions were significantly higher than in healthy controls. Also, MMP-9 in CRC was higher than in non-advanced adenomas. Stool MMP-9 was significantly higher than in healthy controls. Also, MMP-9 was significantly higher than in hea	Gimeno-Gar				-
2016 [47] perplastic polyps, and blood and tissue than in healthy controls. Also, MMP-9 in CRC was higher than in non-advanced adenomas. Patients' stool samples, adenomas, hyperplastic polyps, and perplastic polyps, and single perplastic perplastic perplastic perplastic perplastic perplastic perplastic perplastic perplastic perplasti					-
Patients' stool samples, ade- Annaha'zi A, et al., MMP-9 nomas, hypolyps, and polyps, and polyps, and polyps, and polyps, and compared with all the other polyps, and polyps, and polyps, and plood and tissue than in healthy controls. Also, MMP-9 in CRC was higher than in non-advanced adenomas. Stool MMP-9 was significantly increased in CRC compared with all the other groups. Stool MMP-9 may be a new noninvasive marker in CRC.					
Patients' stool sam- Annaha'zi ples, ade- A, et al., MMP-9 nomas, hy- 2016 [48] perplastic polyps, and higher than in non-advanced adenomas. Stool MMP-9 was significantly increased in CRC compared with all the other groups. Stool MMP-9 may be a new noninvasive marker in CRC	. ,				-
Patients' Stool MMP-9 was significantly increased in CRC compared with all the other polyps, and polyps, and polyps, and adenomas. Stool MMP-9 was significantly increased in CRC compared with all the other groups. Stool MMP-9 may be a new poninyasive marker in CRC compared with all the other groups.					
Patients' stool sam- Annaha'zi ples, ade- A, et al., MMP-9 nomas, hy- 2016 [48] perplastic polyps, and Stool MMP-9 was significantly increased in CRC compared with all the other groups. Stool MMP-9 was significantly increased in CRC compared with all the other groups. Stool MMP-9 may be a new noninvasive marker in CRC					=
Annaha'zi stool sam- A, et al., MMP-9 nomas, hy- 2016 [48] perplastic polyps, and stool sam- perplastic polyps, and stool sam- cantly increased in CRC compared with all the other groups. Stool MMP-9 was significantly increased in CRC sompared with all the other groups. Stool MMP-9 was significantly increased in CRC compared with all the other groups. Stool MMP-9 was significantly increased in CRC compared with all the other groups.			Dation to!		agenomas.
Annaha'zi ples, ade- A, et al., MMP-9 nomas, hy- 2016 [48] perplastic polyps, and cantly increased in CRC compared with all the other groups. Stool Salli- cantly increased in CRC compared with all the other groups. Stool MMP-9 may be a new noninvasive marker in CRC					Stool MMP-9 was signifi-
A, et al., MMP-9 nomas, hy- 2016 [48] perplastic polyps, and ELISA. groups. Stool MMP-9 may be a new noninvasive marker in CRC	A, et al.,	MMP-9			
2016 [48] perplastic groups. Stool MMP-9 may be a new polyps, and popinyasiye marker in CRC			-	FI ISA	compared with all the other
polyps, and noninvasive marker in CRC			•	ELISA.	groups.
noninvasive marker in CKC					Stool MMP-9 may be a new
			CRC tissue		noninvasive marker in CRC.
Expression levels of MMP-7.			Serum specimens of patients with colon adenocarci- noma	Luminex based multiplex assay	Expression levels of MMP-7.
MMP-10 and MMP-12 in	Klupp et al., 2016 [49]				=
MIMP-/, specimens or serum of colon cancer pa-					
Riupp et al., MiviP-10, patients Luminex based multiplex as-					_
2016 [49] and with colon say with serum specimens of the					=
healthy control group. The					=
upregulation of MMP-7,					upregulation of MMP-7,

Cancers **2021**, 13, 6226 7 of 21

				MMP-10, and MMP-12 in
				colon cancer patients' serum
				was associated with a poor
				prognosis.
				The MMP-9 levels were higher in advanced adeno- mas and CRC compared with
Otero-Estévo z O, et al., 2015 [50]	e MMP-9	Human ad- enomas and CRC	non-invasive stool immuno- chemical test (FIT) and ELISA	those reported in samples of healthy individual. Elevated MMP-9 concentra-
				tion was associated with several lesions, size, and adenoma histology.
Bengi G, et	MMP-7,	Human ad-		The expression of <i>TIMP-1</i> , <i>COX-2</i> , and <i>MMP-7</i> was significantly higher in polyps compared with normal tissue.
al., T	TIMP-1, and COX-2	enomas and CRC	Real-time PCR in	Overexpression of MMP-7, COX-2, and TIMP-1 deter- mine an important role of these genes in the progres- sion of colon cancer.
Odabasi M, et al., 2014 [52]	MMP-9 and NGAL	Human ad- enomas and CRC	Immunohistochemistry	The MMP-9 and NGAL overexpression in neoplastic polyps might be used as markers to separate them from non-neoplastic polyps. These genes as immune-histochemical markers determine dysplasia in the early steps of the colorectal adenoma-carcinoma sequence.
Qasim BJ, et al., 2013 [53]	MMP-7	Human ad- enomas	Immunohistochemistry	MMP-7 was expressed in advanced colorectal adeno- matous polyps with large size, severe dysplasia, and villous.
			The MMP enzyme activity	
Sheth RA, et al., 2012 [54]	MIMIP-2,	Xenograft model of CRC in nude mice	was measured by an en- zyme-activatable optical mo- lecular probe and quantitative fluorescence colonoscopy in nude mice which received celecoxib versus vehicle.	There was an apparent linear relationship between meas- ured MMP activity and tu- mor growth rate.
Murname MJ, et al., 2009 [55]	MMP-2 and MMP-9	Mouse models of CRC and human HT-29 CRC cell line	Gene-expression microarray and ELISA	The plotted receiver operating characteristic (ROC) curves estimated the sensitivity and specificity profiles of MMP-2 and MMP-9 for the identification of CRC.
Jeffery N, et al., 2009 [56]	MMP-1, 2, 3, 7, 9, 13, MT1-MMP, MT2-MMP	Human ad- enomas and CRC	Immunohistochemistry	MMP-1, MMP-2, MMP-3, TIMP-1, and TIMP-2 showed a significant increase in car- cinomatous epithelium

Cancers 2021, 13, 6226 8 of 21

	and TIMP-1, TIMP-2, and IMP-3			compared with adenoma epithelium. The increased expression of MMPs and TIMPs occurred at an early stage of colorectal
	The func-			neoplasia.
Lièvre A, et al., 2006 [57]	tional gene promoter polymor- phisms of <i>MMP1</i> , <i>MMP3</i> , and <i>MMP7</i>	Human ad- enomas	Real-time PCR allelic discrimination assay	These data showed a relation between MMP-1 -1607 ins/del G and MMP-3 -1612 ins/del A combined poly- morphisms and risk of small adenomas.
Tutton MG, et al., 2003 [58]	MMP-2 and MMP-9	Patients' plasma samples, adenomas, and CRC	Immunohistochemistry, re- al-time PCR, and ELISA	The expression of MMP-2 and MMP-9 was significantly increased in CRC tissues compared with matched normal tissues. Plasma MMP-2 and MMP-9 levels were significantly elevated at all stages in CRC patients. Plasma levels of these enzymes may be a noninvasive indicator of invasion or metastasis in CRC.

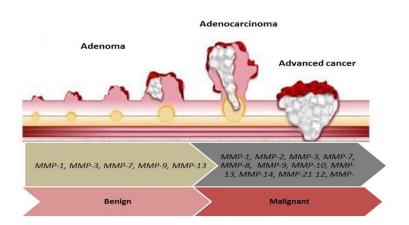


Figure 2. The diagram indicates the role of *MMPs* genes in adenoma development, colorectal adenoma-carcinoma sequence, and tumor progression. *MMP-1, MMP-3, MMP-7, MMP-9,* and *MMP-13* are involved in adenoma development. *MMP-1, MMP-2, MMP-3, MMP-7, MMP-8, MMP-9, MMP-12, MMP-13, MMP-14,* and *MMP-21* participate in adenoma-carcinoma sequence and tumor progression.

Cancers **2021**, 13, 6226 9 of 21

4. The Function of MMPs in Colorectal Polyps and Cancer

4.1. MMP-1, MMP-13, and MMP-8 (Collagenases)

The specific targets for MMP-1 and MMP-13 are in the intestine. MMP-1 can digest type I, II, III, VII, VIII, X collagen, and gelatin. Upregulation of *MMP-1* gene was detected in CRC patients compared to normal tissue [6,59]. Eiro et al. found overexpression of *MMP-1* gene in serrated, villous, and tubulovillous adenomas (i.e., polyps with high potential for transformation to CRC) [44]. Previous investigations demonstrated the correlation between *MMP-1* gene expression and CRC progression: high expression levels of *MMP-1* were associated with invasion, advanced stage metastasis, LNM, and shorter overall survival [60,61]. Wang, et al. investigated the role of MMP-1 in the development of CRC. They found that the downregulation of MMP-1 expression inhibited the progression of CRC in vitro and in vivo by suppressing the PI3K/Akt/c-myc signaling pathway and the EMT [6].

MMP-13, another member of the collagenase category, could degenerate type III collagen. According to the strength of the association between pathologic stage and immunoreactivity scoring (IRS) of MMP-13, in high-grade adenomas and CRC, MMP-13 was observed with a moderate and strong staining intensity, respectively [46]. This result indicated that MMP-13 could help to predict metastatic behavior and prognosis of early-stage cancerous and precancerous colorectal adenoma [46,62]. The study of the association between grade dysplasia and MMP-13 expression in 137 biopsies from patients with cancerous and non-cancerous colorectal adenomas showed that the high expression level of MMP-13 IRS could be helpful to predict metastatic state, prognosis, and recrudescence at an early stage of cancerous and precancerous colorectal adenoma. Moreover, the upregulation of MMP-13 IRS from low to high-grade adenoma was considered an early predictive cancer biomarker [46]. Meanwhile, several studies confirmed that upregulation of MMP-13 was related to advanced CRC and liver metastasis [62-64]. Also, the expression of MMP-13 on the primary tumor cell surface is increased in inflammatory bowel disease. The expression of MMP-13 is closely related to the progression, early relapse, and high mortality of CRC [63,65].

Another member of collagenase enzymes is MMP-8 which is frequently expressed by neutrophils. MMP-8 cleaves many substrates, such as type I, II, and III collagen. This MMP is mainly considered to play a protective role against cancer. However, more recent findings also suggest an oncogenic function of *MMP-8* gene [66,67].

Sirnio et al. found that enhanced-serum MMP-8 level in CRC patients was significantly related to advanced-stage CRC, distant metastasis, lack of MMR, and poor survival. Thus,, they evidenced that MMP-8 is correlated with inflammation and CRC progression [68].

4.2. MMP-2 and MMP-9 (Gelatinase)

MMP-2 and MMP-9, known as gelatinases, can digest type IV collagen and gelatin [69]. Murname et al. showed that MMP-2 protein activity in adenomas with high-grade dysplasia (HGD) was different from adenomas with low-grade dysplasia (LGD). They suggested that the active *MMP-2* gene could predict CRC malignancy risk in patients with adenomatous polyps [70]. Some studies also indicated high expression levels of MMP-9 protein in adenomas with HGD compared to adenomas with LGD and normal tissue. As such, researchers speculated that upregulation of MMP-9 is a primary event in the CRC adenoma-carcinoma sequence [41,71]. High expression levels of MMP-2 protein in CRC tumors compared to normal mucosa have also been reported [41,72]. In addition, a statistically significant relationship between upregulation of *MMP-2* gene with advanced-stage CRC or CRC progression has been observed [41,73–75]. On this basis, *MMP-2* has been suggested as a potential biomarker to detect CRC progression and predict patient survival. Furthermore, overexpression of the *MMP-2* gene was associated with metastasis of lymph nodes and a decrease of cell adhesion in tumors [73].

Cancers **2021**, 13, 6226 10 of 21

Finally, also the upregulation of *MMP-9* gene was associated with the advanced stage of CRC and suggested as a biomarker predictive of poor overall survival [41,76]. Chen et al. indicated that the overexpression of *MMP-9* gene promoted CRC metastasis through the MKK-3/p38/NF-κB pro-oncogenic pathway. Furthermore, they suggested *MMP-9* gene as a potential molecular target for targeted therapy to treat metastatic CRC patients [76].

On the contrary, some investigations reported that MMP-9 gene has a protective role in CRC by stimulating Notch activation resulting in the activation of p21WAF1/Cip1 leading to the suppression of β -catenin [77,78]. In a recent study, although in colitis-associated colon cancer, Walter et al. confirmed this observation by revealing that MMP-9 protein expression was associated with reduced ROS levels, decreased DNA damage, and stimulated mismatch repair pathway [79].

In an interesting study, Wei et al. by analyzing microbiota in tumors obtained by patients with different prognoses, found that the expression of some inflammatory genes, including *MMP-9*, was associated with the abundance of specific bacteria. High levels of *MMP-9* expression were significantly correlated with the high abundance of *B. fragilis* and *F. nucleatum* whereas a high level of *F. prausnitzii* was associated with down-regulation of *MMP-9* [80].

4.3. MMP-3, MMP-10 (Stromelysin)

Another member of MMPs family is MMP-3, or stromelysin-1, which degrades collagen (types II, III, IV, IX, and X), proteoglycans, fibronectin, laminin, and elastin in ECM.

Sipos et al. found a positive association between MMP-3 protein expression and the adenoma–dysplasia–carcinoma sequence. In particular, they reported that high-grade dysplastic sessile adenomatous-stage and early-stage CRC conditions can be differentiated based on the stroma expression of MMP3 [81]. Meaningful positive associations between the protein expression level of MMP-3, invasion, lymph node metastasis, histological type of CRC, and poorly differentiated tumor were reported by Islekel et al. [82]. MMP-3 can activate other MMPs, such as MMP-1, MMP-7, and MMP-9, to promote the progression of tumor initiation [83,84].

MMP-10 also belongs to the stromelysin family. It can digest collagen types II, III, IV, IX, X, proteoglycans, fibronectin, laminin, and elastin. Also, MMP-10 enhances cell growth and invasion in CRC, and its upregulation was found to be associated with poor survival [49,85].

4.4. MMP-7 (Matrilysin)

MMP-7, or matrilysin, digests fibronectin, laminin, type I collagen, and gelatin. It can provide the right condition for vascularization via cleavage of ECM [86]. A major ratio of MMP-7 expression in tumor cells has been reported. Qasim et al. found MMP-7 protein overexpression in villous adenomatous polyps compared to other types of polyps and demonstrated that MMP-7 protein overexpression is an initial event in CRC carcinogenesis that could lead adenomas to CRC [53]. In our laboratory, we observed high expression levels of *MMP-7* and *VEGF-A* mRNA in adenomatous polyps compared to normal tissue. We found that the expression levels of *MMP-7* and *VEGF-A* genes were higher in villous adenoma than in other types of adenomas. Thus, we concluded that the *MMP-7* gene overexpression has a critical role in colorectal adenoma angiogenesis and could be a primary event in the adenoma-carcinoma sequence [45].

MMP-7 gene can enhance tumor growth and metastasis [87]. Also, MMP-7 activates other MMPs, such as proMMP9 and proMMP2 [88] In addition. MMP-7 exerts a wide spectrum of activities not only as an enzyme but also as a signaling molecule. In fact, it has been shown that MMP-7 trans-activates EGFR by releasing the heparin-binding epidermal growth factor (HB-EGF) in CRC cells, with consequent cell proliferation and apoptosis regulation [89,90].

Cancers 2021, 13, 6226 11 of 21

4.5. MMP-12 (Metalloelastase)

MMP-12, or metalloelastase, can digest different substrates. Several studies considered *MMP-12* gene as an anti-metastatic agent [91,92]. Also, it could inhibit angiogenesis by downregulation of *VEGF* and enhancement of the endogenous angiogenesis inhibitor angiostatin. Overall, the role of *MMP-12* in tumor suppression and increase in overall survival has been widely recognized [93–95].

Importantly, Klupp et al. found higher levels of MMP-12 protein expression in sera of CRC patients compared with those of healthy individuals. Also, they suggested an association between MMP-12 protein expression levels and CRC advanced disease and vascular invasion. Furthermore, a significant correlation between the upregulation of MMP-12 expression and poor survival was shown [49].

4.6. MMP-21 (XMMP)

MMP-21 (XMMP) can degenerate aggrecan (cartilage-specific proteoglycan core protein) in the internal region of ECM [96]. Overexpression of MMP-21 protein in CRC compared with normal tissue was shown in many studies [97,98]. Furthermore, significant associations between MMP-21 protein expression and CRC tumor invasion, lymph node metastasis, and distant metastasis were found [97,99]. Wu et al. showed that MMP-21 not only affected CRC progression but also was an independent prognostic biomarker in patients with stage II and stage III CRC cancer. Taken together, these facts led them to conclude that MMP-21 could be used for targeted therapy in CRC [97]. Huang et al. demonstrated that the upregulation of MMP-21 protein was related to shorter overall survival in patients with CRC [98].

4.7. MMP-14 (MT1-MMP)

MMP-14, called MT1-MMP, acts on matrix substrates, such as collagens I, II, III, and gelatin. The *MMP-14* gene plays a crucial role in many biological and pathological conditions and activation of proMMP2 [92,100]. The role of *MMP-14* in angiogenesis and cancer invasion has been identified by previous investigations [101–103]. Cui et al. observed statistically significant associations between the overexpression of *MMP-14* gene in CRC compared to normal mucosa. Their analysis indicated that high expression levels of *MMP-14* were associated with advanced-stage CRC, lymph node metastasis, and poor overall survival. They concluded that the *MMP-14* gene is an oncogene and may represent a potential prognostic biomarker in CRC [104].

Yang et al., showed in an in vivo CRC model that the STAT3 phosphorylation activity and the overexpression of MMP14 protein were enhanced by the overexpression of Hes1 gene. Also, they suggested that Hes1 promoted the invasion of colorectal cancerous cells via the STAT3-MMP14 pathway [103]. It was reported that the overexpression of MMP-14 protein was associated with Prox1 gene. When Prox1 gene was deleted, MMP14 protein was increased, and the mice showed slow-growing, matrix-rich, chemotherapy-resistance, and cancerous cells with malignant stromal features, including activation of fibroblasts, blood vessels dysfunction, and lack of cytotoxic T cells [105].

5. The Effects of Polymorphisms of MMP Genes on Colorectal Carcinogenesis

Single-nucleotide polymorphisms (SNPs) are a common genetic variation involving a single base pair in DNA. SNPs are mostly located in the gene promoter region and may have an impact on gene and protein expression levels. The effects of MMP polymorphisms have been observed in many cancers such as CRC and hepatocellular carcinoma [106,107].

In a Japanese population, the *MMP-1* 1G/2G polymorphism was detected and associated with the development of CRC [108]. In the Iranian population, Kouhkan et al. demonstrated that *MMP-1* 2G/2G genotype polymorphism was correlated with invasion risk of CRC, especially in smoker men [109]. In the Netherlands, *MMP-2* -1306C>T SNP

Cancers **2021**, 13, 6226 12 of 21

was detected in CRC patients, and the T/T genotype was found to be associated with poor overall survival whereas C/C and C/T genotypes showed better outcomes. No difference in overall survival was instead observed among patients with different genotypes of the MMP-9 -1562C>T SNP [110]. Also, in a cohort study of Taiwanese CRC patients, Ting et al. indicated that patients carrying the A/A genotype of the MMP-2-1575G>A SNP had a higher risk to develop distant metastasis compared with patients carrying the T/T genotype [111]. In a Polish population with CRC, individuals with the G/G variant genotype of MMP-7 -181A>G SNP had a higher risk of lymph node involvement and advanced tumor infiltration than patients carrying the A/A genotype [112]. A Chinese study showed that the MMP-9 R279Q SNP relative to the R/R genotype was correlated with a higher risk of CRC compared with the QQ genotype. Also, the allele frequency of the MMP-1 16071G/2G and MMP-7 181 A/G polymorphisms were not associated with CRC [113]. In a Korean population, the homozygous MMP-9-1562C/C genotype was significantly more frequent in CRC cases than in the control group [114]. In Sweden, researchers found that the A/A genotype of MMP-12-82A>G increased the risk of disseminated malignancy in CRC patients while the A/A genotype of MMP-13-82A>G was not correlated to invasion [115].

Lièvre et al. investigated *MMP-3*, *MMP-7*, and *MMP-1* genes promoter polymorphisms in 295 patients with large adenomas and 302 patients with small adenomas. The analysis revealed a significant association between *MMP-3-1612* ins/del A, *MMP-1-1607* ins/del G polymorphism, and small adenomas; also, adenomas were associated with the combined genotype 2G/2G-6A/6A. However, no significant association between *MMP-7* polymorphism and the development of adenomas was found. The authors suggested that only the study *MMP-3* and *MMP-1* gene promoter polymorphisms had potential roles in the development of adenomas from normal colon epithelial cells or in the earliest steps of CRC [57].

Tai et al. showed that *MMP-8* rs11225395 related to the risk of CRC and worst outcomes in a subpopulation of the Han Chinese population. On this basis, they suggested *MMP-8* rs11225395 polymorphism as a potential biomarker predictive of CRC susceptibility [116].

6. Targeting MMPs in CRC Treatment

6.1. Pharmacological Inhibition

Several pharmacological inhibitors of MMPs (MMPIs) have been studied and tested in phase I-III clinical trials, but to date, none of these drugs has been approved for the treatment of cancer, including CRC. Overall, the late stages of the clinical experimentation failed because of the substantial toxicity and weak selectivity of MMPIs [117]. Mainly, candidate MMPIs are represented by small molecules, peptides, and antibodies [118]. Currently, only one broad-spectrum MMPI has been approved by FDA but it has not indication in cancer (i.e., the small molecule periostat) [117,119]. Other MMPIs, such as the small molecule prinomastat, selective for MMP-1, MMP-2, and MMP-9 [120–123] and the GA-5745/andecaliximab, a selective anti-body against MMP-9, have reached the phase III [124,125]. However, none of these trials includes CRC.

6.2. Inhibition of MMPs by TIMPs

Since MMPs are naturally inhibited by TIMPs, these proteins have also been widely investigated mainly to exploit their ability to discover potential strategies for MMP inhibition [126]. The TIMP family consists of four members of proteins (TIMP1-4) that form a 1:1 complex with MMPs. Dysregulation of this complex due to the increased expression of MMPs or a decreased control by TIMPs has been observed in several diseases, including cancer. TIMPs control the activity of MMPs via binding to them (Figure 3) [126–128].

Cancers **2021**, 13, 6226 13 of 21

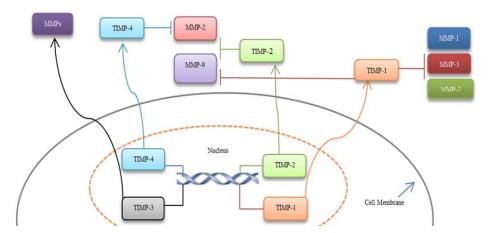


Figure 3. MMPs inhibition by TIMPs. TIMP-1 inhibits MMP-1, 3, 7, 9. TIMP-2 can suppress MMP-2 and 9, and TIMP-4 blocks MMP-2. These inhibitions result in the primary tumor transitioning to advanced CRC. Moreover, TIMP-3 has a protective effect on CRC cases and could bind to several MMPs [126-128].

TIMP-1 inhibits MMP-1, 3, 7, 9 and affects angiogenesis [37,129]. Previous investigations considered a dual activity for the *TIMP-1* gene: in particular, *TIMP-1* was associated with tumor growth at the early stages of colon cancer, and decreased activity of *TIMP-1* could lead to tumor invasion [130,131].

TIMP-2 can suppress MMP-2, MMP-9, and microvascularization [132,129]. Also, downregulation of TIMP-2 is related to invasive CRC [133]. Wang et al. reported that downregulation of *TIMP*-2 in CRC tumor tissues was meaningfully correlated with the depth of invasion, lymph node metastasis, tumor stage, and poor survival [134].

TIMP-3 is known as a tumor suppressor gene and inhibits several MMPs. TIMP-3 downregulation is associated with advanced CRC [135]. Lin et al. represented that, adenovirus-mediated TIMP-3 transduction in CT26 colon cancer cell line suppressed cell growth and stimulated apoptosis. Also, TIMP-3 transduction inhibited migration and invasion. In vivo data indicated that TIMP-3 prevented in vivo tumor growth and liver metastasis [136].

TIMP-4 protein suppresses MMP-2, and one study showed that overexpression of TIMP-4 increased the survival rate of rectal cancer [128].

Currently, no drug mimicking the TIMP activity has been obtained as well as no gene therapeutic approach able to modulate the activity of TIMPs is available.

6.3. MMPs Regulation by microRNA

MicroRNAs, a class of small, endogenous RNAs of 21–25 nucleotides in length, control gene and protein regulation via binding and digesting target mRNA (Table 3). Suppression of MMPs by microRNAs is a suggested way for CRC treatment. Some evidence has been provided. In particular, microRNA-34 (miR-34a) plays a role as a tumor suppressor, and its overexpression could suppress *MMP-1*, *MMP-9*, and tumor cell proliferation, migration, and invasion via acetylation of *P53* in CRC [137–139]. The upregulation of miR-139 reduces proliferation, migration, and invasion by suppression of the IGF-IR/MEK/ERK signaling and *MMP-2* gene in CRC patients [140]. Upregulation of miR-29a increases CRC metastasis via suppression of KLF4 (Kruppel-like factor 4), transcription factor, and upregulation of *MMP-2* gene [141]. Also, miR-29b suppresses CRC metastasis, reduces angiogenesis and EMT by targeting the *MMP-2* gene [142]. Overexpression of miR-143 can suppress the *MMP-7* gene directly and prevent colorectal tumor cell proliferation and invasion [143].

Cancers 2021, 13, 6226 14 of 21

MicroRNA	MMP	Result
miR-34a	MMP-1, MMP-9	miR-34a overexpression prevents tumor cell prolif-
m1K-34a		eration, migration, and invasion [138,139].
miR-139	MMP-2	Downregulation of miR-139 reduces proliferation,
IIIIK-139		migration, and invasion [140].
miR-29a	MMP-2	Upregulation of miR-29a increases metastasis [141].
miR-29b	MMP-2	Upregulation of miR-29b increases metastasis [142].
:D 142	MMP-7	Upregulation of miR-143 enhances tumor cell prolif-
miR- 143		eration and invasion [143].

Table 3. MMPs are Regulated by microRNAs in CRC.

6.4. MMPs Regulation by Long Non-Coding RNAs

Long non-coding RNAs (lncRNAs) can regulate gene expression and have key roles in cell proliferation, migration, invasion, apoptosis, metastasis, and EMT in CRC. In this regard, lncRNA-targeted therapy is today considered a potential promising strategy for CRC treatment [144]. In fact, based on mechanistic studies investigating the complex lncRNA-mediated sponge interactions in CRC, potential therapeutic targets for the treatment of this cancer may be identified. Among the available findings, Tian et al. demonstrated that the suppression of TUG1 by shRNA prevented MMP-14 expression, proliferation, invasion, and EMT in colon cancer [145]. Sun et al. found a significant association between XIST inhibition and suppression of c-Myc, cyclinD1, and MMP-7 expression through inactivation of Wnt/β-catenin signaling pathway [146]. A recent investigation showed a meaningful correlation between the overexpression of LINC00963 and the upregulation of MMP-2 and MMP-9, proliferation, migration, and invasion of CRC cells [147]. Duan et al. revealed that the inhibition of the CCEPR lncRNA reduced the expression levels of MMP-2 and MMP-9, and prevented EMT in CRC cells [148]. Pan et al. realized that the expression level of MMP-2 protein was notably decreased when PCA3 was knocked out. In addition, suppression of PCA3 inhibited colon cancer cell invasion and migration [149].

7. Conclusions

In summary, MMPs genes and proteins, through complex mechanisms involving the induction of many molecular signaling pathways and the EMT process, play a relevant role in the transition from pre-cancerous lesions and polyps to advanced CRC. However, further investigation is needed to understand how MMPs exactly work. This would improve the selectivity of MMPIs that could be exploited in a dual-mode: to treat CRC alone or in combination with targeted agents and/or chemotherapy and to prevent CRC development.

Author Contributions: Conceptualization, investigation, writing—original draft, designed tables and figures Z.P.; editing, validation, and revise, S.N.; writing, design table, and investigation, N.P.; revise, B.S.; investigation, H.N. and H.S.; validation, H.A.-A. and E.M.; supervision, validation, and revise E.N.-M. and M.A.B. All authors have read and agreed to the published version of the manuscript.

Funding: This research was supported by Fondo Beneficenza Intesa Sanpaolo S.p.A. (Milan, Italy) and Associazione Giacomo Onlus (Castiglioncello, Italy) to E.M.

Acknowledgments: The authors would like to thank all the staff of the Department of Cancer at the Research Institute for Gastroenterology and Liver Diseases, Shahid Beheshti University of Medical Sciences, Tehran, Iran.

Conflicts of Interest: The authors declare no conflict of interests.

Cancers **2021**, 13, 6226 15 of 21

References

1. Sung, H.; Ferlay, J.; Siegel, R.L.; Laversanne, M.; Soerjomataram, I.; Jemal, A.; Bray, F. Global Cancer Statistics 2020: GLO-BOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA Cancer J. Clin.* 2021, 71, 209–249, doi:10.3322/caac.21660.

- 2. Vogelstein, B.; Fearon, E.R.; Hamilton, S.R.; Kern, S.E.; Preisinger, A.C.; Leppert, M.; Nakamura, Y.; White, R.; Smits, A.M.; Bos, J.L. Genetic alterations during colorectal-tumor development. N. Engl. J. Med. 1988, 319, 525–532, doi:10.1056/nejm198809013190901.
- 3. Kasi, A.; Handa, S.; Bhatti, S.; Umar, S.; Bansal, A.; Sun, W. Molecular Pathogenesis and Classification of Colorectal Carcinoma. *Curr. Colorectal Cancer Rep.* **2020**, *16*, 97–106, doi:10.1007/s11888-020-00458-z.
- Huck, M.B.; Bohl, J.L. Colonic Polyps: Diagnosis and Surveillance. Clin. Colon Rectal Surg. 2016, 29, 296–305, doi:10.1055/s-0036-1584091.
- 5. Smit, W.L.; Spaan, C.N.; de Boer, R.J.; Ramesh, P.; Garcia, T.M.; Meijer, B.J.; Vermeulen, J.L.M.; Lezzerini, M.; MacInnes, A.W.; Koster, J.; et al. Driver mutations of the adenoma-carcinoma sequence govern the intestinal epithelial global translational capacity. *Proc. Natl. Acad. Sci. USA* **2020**, *117*, 25560, doi:10.1073/pnas.1912772117.
- 6. Wang, K.; Zheng, J.; Yu, J.; Wu, Y.; Guo, J.; Xu, Z.; Sun, X. Knockdown of MMP--1 inhibits the progression of colorectal cancer by suppressing the PI3K/Akt/c--myc signaling pathway and EMT. *Oncol. Rep.* **2020**, *43*, 1103–1112, doi:10.3892/or.2020.7490.
- 7. Yuan, X.; Sun, X.; Shi, X.; Wang, H.; Wu, G.; Jiang, C.; Yu, D.; Zhang, W.; Xue, B.; Ding, Y. USP39 promotes colorectal cancer growth and metastasis through the Wnt/β-catenin pathway. *Oncol. Rep.* **2017**, *37*, 2398–2404, doi:10.3892/or.2017.5454.
- 8. Quintero-Fabián, S.; Arreola, R.; Becerril-Villanueva, E.; Torres-Romero, J.C.; Arana-Argáez, V.; Lara-Riegos, J.; Ramí-rez-Camacho, M.A.; Alvarez-Sánchez, M.E. Role of Matrix Metalloproteinases in Angiogenesis and Cancer. *Front. Oncol.* **2019**, 9, 1370, doi:10.3389/fonc.2019.01370.
- 9. Yadav, L.; Puri, N.; Rastogi, V.; Satpute, P.; Ahmad, R.; Kaur, G. Role of Matrix Metalloproteinases in Angiogenesis and Cancer. *Asian Pac. J. Cancer Prev.* **2014**, *15*, 1085–1091, doi:10.7314/apjcp.2014.15.3.1085.
- 10. Ligi, D.; Mannello, F. Do matrix metalloproteinases represent reliable circulating biomarkers in colorectal cancer? *Br. J. Cancer* **2016**, *115*, 633–634, doi:10.1038/bjc.2016.241.
- 11. Yeh, Y.-C.; Sheu, B.-S. Matrix metalloproteinases and their inhibitors in the gastrointestinal cancers: Current knowledge and clinical potential. *Met. Med.* **2014**, *1*, 3–13, doi:10.2147/MNM.S51000.
- 12. Wu, Z.; Liu, Z.; Ge, W.; Shou, J.; You, L.; Pan, H.; Han, W. Analysis of potential genes and pathways associated with the colorectal normal mucosa-adenoma-carcinoma sequence. *Cancer Med.* **2018**, *7*, 2555–2566, doi:10.1002/cam4.1484.
- Bertelson, N.L.; Kalkbrenner, K.A.; Merchea, A.; Dozois, E.J.; Landmann, R.G.; De Petris, G.; Young-Fadok, T.M.; Etzioni, D.A. Colectomy for Endoscopically Unresectable Polyps: How Often Is It Cancer? *Dis. Colon Rectum* 2012, 55, 1111–1116, doi:10.1097/DCR.0b013e3182695115.
- 14. Shussman, N.; Wexner, S.D. Colorectal polyps and polyposis syndromes. *Gastroenterol. Rep.* **2014**, 2, 1–15, doi:10.1093/gastro/got041.
- 15. Liljegren, A.; Lindblom, A.; Rotstein, S.; Nilsson, B.; Rubio, C.; Jaramillo, E. Prevalence and incidence of hyperplastic polyps and adenomas in familial colorectal cancer: Correlation between the two types of colon polyps. *Gut* 2003, 52, 1140–1147, doi:10.1136/gut.52.8.1140.
- 16. Malki, A.; ElRuz, R.A.; Gupta, I.; Allouch, A.; Vranic, S.; Al Moustafa, A.E. Molecular Mechanisms of Colon Cancer Progression and Metastasis: Recent Insights and Advancements. *Int. J. Mol. Sci.* **2020**, 22, 130, doi:10.3390/ijms22010130.
- 17. Kato, S.; Lippman, S.M.; Flaherty, K.T.; Kurzrock, R. The Conundrum of Genetic "Drivers" in Benign Conditions. *J. Natl. Cancer Inst.* **2016**, *108*, djw036, doi:10.1093/jnci/djw036.
- 18. Mustjoki, S.; Young, N.S. Somatic Mutations in "Benign" Disease. N. Engl. J. Med. **2021**, 384, 2039–2052, doi:10.1056/NEJMra2101920.
- Nojadeh, J.N.; Behrouz Sharif, S.; Sakhinia, E. Microsatellite instability in colorectal cancer. EXCLI J. 2018, 17, 159–168, doi:10.17179/excli2017-948.
- 20. Arabsorkhi, Z.; Sadeghi, H.; Gharib, E.; Rejali, L.; Asadzadeh-Aghdaei, H.; Nazemalhosseini-Mojarad, E. Can *hypoxia-inducible factor-1α* overexpression discriminate human colorectal cancers with different microsatellite instability? *Genes Genet. Syst.* **2021**, 96, 1-6, doi:10.1266/ggs.21-00026.
- Pino, M.S.; Chung, D.C. The chromosomal instability pathway in colon cancer. Gastroenterology 2010, 138, 2059–2072, doi:10.1053/j.gastro.2009.12.065.
- 22. Colussi, D.; Brandi, G.; Bazzoli, F.; Ricciardiello, L. Molecular pathways involved in colorectal cancer: Implications for disease behavior and prevention. *Int. J. Mol. Sci.* **2013**, *14*, 16365–16385, doi:10.3390/ijms140816365.
- 23. Grady, W.M.; Markowitz, S.D. The molecular pathogenesis of colorectal cancer and its potential application to colorectal cancer screening. *Dig. Dis. Sci.* **2015**, *60*, 762–772, doi:10.1007/s10620-014-3444-4.
- 24. Sadanandam, A.; Lyssiotis, C.A.; Homicsko, K.; Collisson, E.A.; Gibb, W.J.; Wullschleger, S.; Ostos, L.C.; Lannon, W.A.; Grotzinger, C.; Del Rio, M.; et al. A colorectal cancer classification system that associates cellular phenotype and responses to therapy. *Nat. Med.* **2013**, *19*, 619–625, doi:10.1038/nm.3175.
- 25. Guinney, J.; Dienstmann, R.; Wang, X.; de Reyniès, A.; Schlicker, A.; Soneson, C.; Marisa, L.; Roepman, P.; Nyamundanda, G.; Angelino, P.; et al. The consensus molecular subtypes of colorectal cancer. *Nat. Med.* **2015**, *21*, 1350–1356, doi:10.1038/nm.3967.

Cancers **2021**, 13, 6226 16 of 21

26. Thanki, K.; Nicholls, M.E.; Gajjar, A.; Senagore, A.J.; Qiu, S.; Szabo, C.; Hellmich, M.R.; Chao, C. Consensus Molecular Subtypes of Colorectal Cancer and their Clinical Implications. *Int. Biol. Biomed. J.* **2017**, *3*, 105–111.

- 27. McCorry, A.M.; Loughrey, M.B.; Longley, D.B.; Lawler, M.; Dunne, P.D. Epithelial-to-mesenchymal transition signature assessment in colorectal cancer quantifies tumour stromal content rather than true transition. *J. Pathol.* **2018**, 246, 422–426, doi:10.1002/path.5155.
- 28. Isella, C.; Brundu, F.; Bellomo, S.E.; Galimi, F.; Zanella, E.; Porporato, R.; Petti, C.; Fiori, A.; Orzan, F.; Senetta, R.; et al. Selective analysis of cancer-cell intrinsic transcriptional traits defines novel clinically relevant subtypes of colorectal cancer. *Nat. Commun.* 2017, *8*, 15107, doi:10.1038/ncomms15107.
- **29.** Cui, N.; Hu, M.; Khalil, R.A. Biochemical and biological attributes of matrix metalloproteinases. *Prog. Mol. Biol. Transl. Sci.* **2017**, 147, 1–73, doi:10.1016/bs.pmbts.2017.02.005.
- 30. Morgunova, E.; Tuuttila, A.; Bergmann, U.; Isupov, M.; Lindqvist, Y.; Schneider, G.; Tryggvason, K. Structure of human pro-matrix metalloproteinase-2: Activation mechanism revealed. *Science* **1999**, 284, 1667–1670, doi:10.1126/science.284.5420.1667.
- 31. Laronha, H.; Caldeira, J. Structure and function of human matrix metalloproteinases. *Cells* **2020**, *9*, 1076, doi:10.3390/cells9051076.
- 32. Sternlicht, M.D.; Werb, Z. How matrix metalloproteinases regulate cell behavior. *Annu. Rev. Cell. Dev. Biol.* **2001**, *17*, 463–516, doi:10.1146/annurev.cellbio.17.1.463.
- 33. Baker, A.H.; Edwards, D.R.; Murphy, G. Metalloproteinase inhibitors: Biological actions and therapeutic opportunities. *J. Cell Sci.* 2002, 115, 3719–3727, doi:10.1242/jcs.00063.
- 34. Hua, H.; Li, M.; Luo, T.; Yin, Y.; Jiang, Y. Matrix metalloproteinases in tumorigenesis: An evolving paradigm. *Cell Mol. Life Sci.* **2011**, *68*, 3853–3868, doi:10.1007/s00018-011-0763-x.
- 35. Alaseem, A.; Alhazzani, K.; Dondapati, P.; Alobid, S.; Bishayee, A.; Rathinavelu, A. Matrix Metalloproteinases: A challenging paradigm of cancer management. *Semin. Cancer Biol.* **2019**, *56*, 100–115, doi:10.1016/j.semcancer.2017.11.008.
- 36. Gonzalez-Avila, G.; Sommer, B.; Mendoza-Posada, D.A.; Ramos, C.; Garcia-Hernandez, A.A.; Falfan-Valencia, R. Matrix metalloproteinases participation in the metastatic process and their diagnostic and therapeutic applications in cancer. *Crit. Rev. Oncol. Hematol.* **2019**, 137, 57–83, doi:10.1016/j.critrevonc.2019.02.010.
- 37. Herszényi, L.; Hritz, I.; Lakatos, G.; Varga, M.Z.; Tulassay, Z. The behavior of matrix metalloproteinases and their inhibitors in colorectal cancer. *Int. J. Mol. Sci.* **2012**, *13*, 13240–13263, doi:10.3390/ijms131013240.
- 38. Huang, X.; Lan, Y.; Li, E.; Li, J.; Deng, Q.; Deng, X. Diagnostic values of MMP-7, MMP-9, MMP-11, TIMP-1, TIMP-2, CEA, and CA19-9 in patients with colorectal cancer. *J. Int. Med. Res.* **2021**, *49*, 1–11, doi:10.1177/03000605211012570.
- 39. Zhou, X.; Yang, D.; Ding, X.; Xu, P. Clinical value of microRNA-135a and MMP-13 in colon cancer. *Oncol. Lett.* **2021**, 22, 583, doi:10.3892/ol.2021.12844.
- 40. Rasool, M.; Malik, A.; Waquar, S.; Ain, Q.T.; Rasool, R.; Asif, M.; Anfinan, N.; Haque, A.; Alam, H.; Ahmed, S.; et al. Assessment of clinical variables as predictive markers in the development and progression of colorectal cancer. *Bioengineered* **2021**, *12*, 2288–2298, doi:10.1080/21655979.2021.1933680.
- 41. Barabás, L.; Hritz, I.; István, G.; Tulassay, Z.; Herszényi, L. The Behavior of MMP-2, MMP-7, MMP-9, and Their Inhibitors TIMP-1 and TIMP-2 in Adenoma-Colorectal Cancer Sequence. *Dig. Dis.* **2021**, *39*, 217–224, doi:10.1159/000511765.
- 42. Hsieh, S.L.; Hsieh, S.; Lai, P.Y.; Wang, J.J.; Li, C.C.; Wu, C.C. Carnosine Suppresses Human Colorectal Cell Migration and Intravasation by Regulating EMT and MMP Expression. *Am. J. Chin. Med.* **2019**, *47*, 477–494, doi:10.1142/S0192415X19500241.
- 43. Kıyak, R.; Keles, D.; Bengi, G.; Yalcin, M.; Topalak, Ö.; Oktay, G. The Importance of Fecal and Plasma CEA, COX-2, MMP-7, and TIMP-1 in the Diagnosis of Colorectal Cancer. *J. Basic Clin. Health Sci.* **2018**, 2, 7–14, doi:10.30621/jbachs.2018.259.
- 44. Eiró, N.; Gonzalez, L.; Cid, S.; Andicoechea, A.; Vizoso, F. Matrix metalloproteases expression in different histological types of colorectal polyps. *Rev. Esp. Enferm. Dig.* **2017**, 109, 414–420, doi:10.17235/reed.2017.4551/2016.
- 45. Pezeshkian, Z.; Forouzesh, F.; Peyravian, N.; Yaghoob-Taleghani, M.; Asadzadeh-Aghdaei, H.; Zali, M.; Nazemalhosseini-Mojarad, E. Clinicopathological correlations of *VEGF-A* and *MMP-7* genes expression in different types of colorectal adenoma polyps. *WCRJ* **2017**, *4*, e978.
- 46. Wernicke, A.-K.; Churin, Y.; Sheridan, D.; Windhorst, A.; Tschuschner, A.; Gattenlöhner, S.; Roderfeld, M.; Roeb, E. Matrix metalloproteinase-13 refines pathological staging of precancerous colorectal lesions. *Oncotarget* **2016**, *7*, 73552–73557, doi:10.18632/oncotarget.12429.
- 47. Gimeno-García, A.Z.; Triñanes, J.; Quintero, E.; Salido, E.; Nicolás-Pérez, D.; Adrián-de-Ganzo, Z.; Alarcón-Fernández, O.; Abrante, B.; Romero, R.; Carrillo, M.; et al. Plasma matrix metalloproteinase 9 as an early surrogate biomarker of advanced colorectal neoplasia. *Gastroenterol. Hepatol.* **2016**, *39*, 433–441, doi:10.1016/j.gastrohep.2015.10.002.
- 48. Annaházi, A.; Ábrahám, S.; Farkas, K.; Rosztóczy, A.; Inczefi, O.; Földesi, I.; Szűcs, M.; Rutka, M.; Theodorou, V.; Eutamene, H.; et al. A pilot study on faecal MMP-9: A new noninvasive diagnostic marker of colorectal cancer. *Br. J. Cancer* **2016**, *114*, 787–792, doi:10.1038/bjc.2016.31.
- 49. Klupp, F.; Neumann, L.; Kahlert, C.; Diers, J.; Halama, N.; Franz, C.; Schmidt, T.; Koch, M.; Weitz, J.; Schneider, M.; et al. Serum MMP7, MMP10 and MMP12 level as negative prognostic markers in colon cancer patients. *BMC Cancer* **2016**, *16*, 494, doi:10.1186/s12885-016-2515-7.

Cancers **2021**, 13, 6226 17 of 21

Otero-Estévez, O.; De Chiara, L.; Rodríguez-Girondo, M.; Rodríguez-Berrocal, F.J.; Cubiella, J.; Castro, I.; Hernández, V.; Martínez-Zorzano, V.S. Serum matrix metalloproteinase-9 in colorectal cancer family-risk population screening. Sci. Rep. 2015, 5, 13030, doi:10.1038/srep13030.

- 51. Bengi, G.; Keles, D.; Topalak, Ö.; Yalçin, M.; Kiyak, R.; Oktay, G. Expressions of TIMP-1, COX-2 and MMP-7 in Colon Polyp and Colon Cancer. *Euroasian J. Hepatogastroenterol.* **2015**, *5*, 74–79, doi:10.5005/jp-journals-10018-1138.
- 52. Odabasi, M.; Yesil, A.; Ozkara, S.; Paker, N.; Ozkan, S.; Eris, C.; Yildiz, M.K.; Abuoglu, H.H.; Gunay, E.; Tekeşin, K. Role of human neutrophil gelatinase associated lipocalin (NGAL) and Matrix Metalloproteinase-9 (MMP-9) overexpression in neoplastic colon polyps. *Int. J. Clin. Exp. Med.* **2014**, *7*, 2804–2811.
- 53. Qasim, B.J.; Ali, H.H.; Hussein, A.G. Immunohistochemical expression of matrix metalloproteinase-7 in human colorectal adenomas using specified automated cellular image analysis system: A clinicopathological study. *Saudi J. Gastroenterol.* **2013**, *19*, 23–27, doi:10.4103/1319-3767.105916.
- 54. Sheth, R.A.; Kunin, A.; Stangenberg, L.; Sinnamon, M.; Hung, K.E.; Kucherlapati, R.; Mahmood, U. In vivo optical molecular imaging of matrix metalloproteinase activity following celecoxib therapy for colorectal cancer. *Mol. Imaging* **2012**, *11*, 417–425.
- 55. Murnane, M.J.; Cai, J.; Shuja, S.; McAneny, D.; Klepeis, V.; Willett, J.B. Active MMP-2 effectively identifies the presence of colorectal cancer. *Int. J. Cancer* **2009**, *125*, 2893–2902, doi:10.1002/ijc.24682.
- 56. Jeffery, N.; McLean, M.H.; El-Omar, E.M.; Murray, G.I. The matrix metalloproteinase/tissue inhibitor of matrix metalloproteinase profile in colorectal polyp cancers. *Histopathology* **2009**, *54*, 820–828, doi:10.1111/j.1365-2559.2009.03301.x.
- 57. Lièvre, A.; Milet, J.; Carayol, J.; Le Corre, D.; Milan, C.; Pariente, A.; Nalet, B.; Lafon, J.; Faivre, J.; Bonithon-Kopp, C.; et al. Genetic polymorphisms of MMP1, MMP3 and MMP7 gene promoter and risk of colorectal adenoma. *BMC Cancer* 2006, 6, 270, doi:10.1186/1471-2407-6-270.
- 58. Tutton, M.G.; George, M.L.; Eccles, S.A.; Burton, S.; Swift, R.I.; Abulafi, A.M. Use of plasma MMP-2 and MMP-9 levels as a surrogate for tumour expression in colorectal cancer patients. *Int. J. Cancer* **2003**, *107*, 541–550, doi:10.1002/ijc.11436.
- 59. Jonsson, A.; Falk, P.; Angenete, E.; Hjalmarsson, C.; Ivarsson, M.-L. Plasma MMP-1 Expression as a Prognostic Factor in Colon Cancer. *J. Surg. Res.* **2021**, *266*, 254–260, doi:10.1016/j.jss.2021.04.021.
- 60. Liang, Y.; Lv, Z.; Huang, G.; Qin, J.; Li, H.; Nong, F.; Wen, B. Prognostic significance of abnormal matrix collagen remodeling in colorectal cancer based on histologic and bioinformatics analysis. *Oncol. Rep.* **2020**, *44*, 1671–1685, doi:10.3892/or.2020.7729.
- 61. Sunami, E.; Tsuno, N.; Osada, T.; Saito, S.; Kitayama, J.; Tomozawa, S.; Tsuruo, T.; Shibata, Y.; Muto, T.; Nagawa, H. MMP-1 is a prognostic marker for hematogenous metastasis of colorectal cancer. *Oncologist* **2000**, *5*, 108–114, doi:10.1634/theoncologist.5-2-108.
- 62. Yamada, T.; Oshima, T.; Yoshihara, K.; Tamura, S.; Kanazawa, A.; Inagaki, D.; Yamamoto, N.; Sato, T.; Fujii, S.; Numata, K.; et al. Overexpression of *MMP-13* gene in colorectal cancer with liver metastasis. *Anticancer Res.* **2010**, *30*, 2693–2699.
- 63. Leeman, M.F.; McKay, J.A.; Murray, G.I. Matrix metalloproteinase 13 activity is associated with poor prognosis in colorectal cancer. J. Clin. Pathol. 2002, 55, 758–762, doi:10.1136/jcp.55.10.758.
- 64. Merchant, N.; Chalikonda, G.; Nagaraju, G.P. Role of Matrix Metalloproteinases in Colorectal Cancer. In *Theranostics Approaches to Gastric and Colon Cancer*; Springer: Singapore, 2020; pp. 49–59, doi:10.1007/978-981-15-2017-4_3.
- 65. Yan, Q.; Yuan, Y.; Yankui, L.; Jingjie, F.; Linfang, J.; Yong, P.; Dong, H.; Xiaowei, Q. The Expression and Significance of CXCR5 and MMP-13 in Colorectal Cancer. *Cell Biochem. Biophys.* **2015**, 73, 253–259, doi:10.1007/s12013-015-0624-6.
- 66. Korpi, J.T.; Kervinen, V.; Mäklin, H.; Väänänen, A.; Lahtinen, M.; Läärä, E.; Ristimäki, A.; Thomas, G.; Ylipalosaari, M.; Aström, P.; et al. Collagenase-2 (matrix metalloproteinase-8) plays a protective role in tongue cancer. *Br. J. Cancer* **2008**, *98*, 766–775, doi:10.1038/sj.bjc.6604239.
- 67. Balbín, M.; Fueyo, A.; Tester, A.M.; Pendás, A.M.; Pitiot, A.S.; Astudillo, A.; Overall, C.M.; Shapiro, S.D.; López-Otín, C. Loss of collagenase-2 confers increased skin tumor susceptibility to male mice. *Nat. Genet.* **2003**, *35*, 252–257, doi:10.1038/ng1249.
- 68. Sirniö, P.; Tuomisto, A.; Tervahartiala, T.; Sorsa, T.; Klintrup, K.; Karhu, T.; Herzig, K.-H.; Mäkelä, J.; Karttunen, T.J.; Salo, T.; et al. High-serum MMP-8 levels are associated with decreased survival and systemic inflammation in colorectal cancer. *Br. J. Cancer* 2018, 119, 213–219, doi:10.1038/s41416-018-0136-4.
- 69. Beutel, B.; Song, J.; Konken, C.P.; Korpos, E.; Schinor, B.; Gerwien, H.; Vidyadharan, R.; Burmeister, M.; Li, L.; Haufe, G.; et al. New in Vivo Compatible Matrix Metalloproteinase (MMP)-2 and MMP-9 Inhibitors. *Bioconjugate Chem.* 2018, 29, 3715–3725, doi:10.1021/acs.bioconjchem.8b00618.
- 70. Murnane, M.J.; Cai, J.; Shuja, S.; McAneny, D.; Willett, J.B. Active matrix metalloproteinase-2 activity discriminates colonic mucosa, adenomas with and without high-grade dysplasia, and cancers. *Hum. Pathol.* **2011**, 42, 688–701, doi:10.1016/j.humpath.2010.08.021.
- 71. Gimeno-García, A.Z.; Santana-Rodríguez, A.; Jiménez, A.; Parra-Blanco, A.; Nicolás-Pérez, D.; Paz-Cabrera, C.; Díaz-González, F.; Medina, C.; Díaz-Flores, L.; Quintero, E. Up-regulation of gelatinases in the colorectal adenoma-carcinoma sequence. *Eur. J. Cancer* 2006, 42, 3246–3252, doi:10.1016/j.ejca.2006.06.025.
- 72. Salem, N.; Kamal, I.; Al-Maghrabi, J.; Abuzenadah, A.; Peer-Zada, A.A.; Qari, Y.; Al-Ahwal, M.; Al-Qahtani, M.; Buhmeida, A. High expression of matrix metalloproteinases: MMP-2 and MMP-9 predicts poor survival outcome in colorectal carcinoma. *Future Oncol.* **2016**, *12*, 323–331, doi:10.2217/fon.15.325.
- 73. Langenskiöld, M.; Holmdahl, L.; Falk, P.; Ivarsson, M.-L. Increased plasma MMP-2 protein expression in lymph node-positive patients with colorectal cancer. *Int. J. Colorectal Dis.* **2005**, 20, 245–252, doi:10.1007/s00384-004-0667-4.

Cancers **2021**, 13, 6226 18 of 21

74. Kryczka, J.; Stasiak, M.; Dziki, L.; Mik, M.; Dziki, A.; Cierniewski, C.S. Matrix metalloproteinase-2 cleavage of the β1 integrin ectodomain facilitates colon cancer cell motility. *J. Biol. Chem.* **2012**, *287*, 36556–36566, doi:10.1074/jbc.M112.384909.

- 75. Takeuchi, T.; Hisanaga, M.; Nagao, M.; Ikeda, N.; Fujii, H.; Koyama, F.; Mukogawa, T.; Matsumoto, H.; Kondo, S.; Takahashi, C.; et al. The membrane-anchored matrix metalloproteinase (MMP) regulator RECK in combination with MMP-9 serves as an informative prognostic indicator for colorectal cancer. *Clin. Cancer Res.* **2004**, *10*, 5572–5579, doi:10.1158/1078-0432.CCR-03-0656.
- 76. Chen, H.; Ye, Y.; Yang, Y.; Zhong, M.; Gu, L.; Han, Z.; Qiu, J.; Liu, Z.; Qiu, X.; Zhuang, G. TIPE-mediated up-regulation of MMP-9 promotes colorectal cancer invasion and metastasis through MKK-3/p38/NF-κB pro-oncogenic signaling pathway. *Signal Transduct. Target. Ther.* **2020**, *5*, 163, doi:10.1038/s41392-020-00276-7.
- 77. Garg, P.; Jeppsson, S.; Dalmasso, G.; Ghaleb, A.M.; McConnell, B.B.; Yang, V.W.; Gewirtz, A.T.; Merlin, D.; Sitaraman, S.V. Notch1 regulates the effects of matrix metalloproteinase-9 on colitis-associated cancer in mice. *Gastroenterology* **2011**, *141*, 1381–1392, doi:10.1053/j.gastro.2011.06.056.
- 78. Garg, P.; Sarma, D.; Jeppsson, S.; Patel, N.R.; Gewirtz, A.T.; Merlin, D.; Sitaraman, S.V. Matrix metalloproteinase-9 functions as a tumor suppressor in colitis-associated cancer. *Cancer Res.* **2010**, *70*, 792–801, doi:10.1158/0008-5472.CAN-09-3166.
- 79. Walter, L.; Canup, B.; Pujada, A.; Bui, T.A.; Arbasi, B.; Laroui, H.; Merlin, D.; Garg, P. Matrix metalloproteinase 9 (MMP9) limits reactive oxygen species (ROS) accumulation and DNA damage in colitis-associated cancer. *Cell Death Dis.* **2020**, *11*, 767, doi:10.1038/s41419-020-02959-z.
- 80. Wei, Z.; Cao, S.; Liu, S.; Yao, Z.; Sun, T.; Li, Y.; Li, J.; Zhang, D.; Zhou, Y. Could gut microbiota serve as prognostic biomarker associated with colorectal cancer patients' survival? A pilot study on relevant mechanism. *Oncotarget* 2016, 7, 46158–46172, doi:10.18632/oncotarget.10064.
- 81. Sipos, F.; Germann, T.M.; Wichmann, B.; Galamb, O.; Spisák, S.; Krenács, T.; Tulassay, Z.; Molnár, B.; Műzes, G. MMP3 and CXCL1 are potent stromal protein markers of dysplasia-carcinoma transition in sporadic colorectal cancer. *Eur. J. Cancer Prev.* **2014**, 23, 336–343, doi:10.1097/CEJ.0000000000000058.
- 82. Işlekel, H.; Oktay, G.; Terzi, C.; Canda, A.E.; Füzün, M.; Küpelioğlu, A. Matrix metalloproteinase-9,-3 and tissue inhibitor of matrix metalloproteinase-1 in colorectal cancer: Relationship to clinicopathological variables. *Cell Biochem. Funct.* **2007**, 25, 433–441, doi:10.1002/cbf.1325.
- 83. Jin, X.; Yagi, M.; Akiyama, N.; Hirosaki, T.; Higashi, S.; Lin, C.Y.; Dickson, R.B.; Kitamura, H.; Miyazaki, K. Matriptase activates stromelysin (MMP-3) and promotes tumor growth and angiogenesis. *Cancer Sci.* **2006**, *97*, 1327–1334, doi:10.1111/j.1349-7006.2006.00328.x.
- 84. Inuzuka, K.; Ogata, Y.; Nagase, H.; Shirouzu, K. Significance of coexpression of urokinase-type plasminogen activator, and matrix metalloproteinase 3 (stromelysin) and 9 (gelatinase B) in colorectal carcinoma. *J. Surg. Res.* **2000**, *93*, 211–218, doi:10.1006/jsre.2000.5952.
- 85. Batra, J.; Robinson, J.; Soares, A.S.; Fields, A.P.; Radisky, D.C.; Radisky, E.S. Matrix metalloproteinase-10 (MMP-10) interaction with tissue inhibitors of metalloproteinases TIMP-1 and TIMP-2: Binding studies and crystal structure. *J. Biol. Chem.* **2012**, 287, 15935–15946, doi:10.1074/jbc.M112.341156.
- 86. Surlin, V.; Ioana, M.; Pleşea, I.E. Genetic patterns of metalloproteinases and their tissular inhibitors—Clinicopathologic and prognostic significance in colorectal cancer. *Rom. J. Morphol. Embryol.* **2011**, *52*, 231–236.
- 87. Asadzadeh Aghdaei, H.; Pezeshkian, Z.; Abdollahpour-Alitappeh, M.; Nazemalhosseini Mojarad, E.; Zali, M.R. The Role of Angiogenesis in Colorectal Polyps and Cancer, a Review. *Med. Lab. J.* **2018**, *12*, 1–6, doi:10.29252/mlj.12.4.1.
- 88. Ii, M.; Yamamoto, H.; Adachi, Y.; Maruyama, Y.; Shinomura, Y. Role of matrix metalloproteinase-7 (matrilysin) in human cancer invasion, apoptosis, growth, and angiogenesis. *Exp. Biol. Med.* **2006**, *231*, 20–27, doi:10.1177/153537020623100103.
- 89. Cheng, K.; Xie, G.; Raufman, J.P. Matrix metalloproteinase-7-catalyzed release of HB-EGF mediates deoxycholyltaurine-induced proliferation of a human colon cancer cell line. *Biochem. Pharmacol.* **2007**, 73, 1001–1012, doi:10.1016/j.bcp.2006.11.028.
- 90. Xie, G.; Cheng, K.; Shant, J.; Raufman, J.P. Acetylcholine-induced activation of M3 muscarinic receptors stimulates robust matrix metalloproteinase gene expression in human colon cancer cells. Am. J. Physiol. Gastrointest. Liver Physiol. 2009, 296, G755–G763, doi:10.1152/ajpgi.90519.2008.
- 91. Decock, J.; Thirkettle, S.; Wagstaff, L.; Edwards, D.R. Matrix metalloproteinases: Protective roles in cancer. *J. Cell Mol. Med.* **2011**, *15*, 1254–1265, doi:10.1111/j.1582-4934.2011.01302.x.
- 92. Asano, T.; Tada, M.; Cheng, S.; Takemoto, N.; Kuramae, T.; Abe, M.; Takahashi, O.; Miyamoto, M.; Hamada, J.; Moriuchi, T.; et al. Prognostic values of matrix metalloproteinase family expression in human colorectal carcinoma. *J. Surg. Res.* **2008**, *146*, 32–42, doi:10.1016/j.jss.2007.02.011.
- 93. Yang, W.; Arii, S.; Gorrin-Rivas, M.J.; Mori, A.; Onodera, H.; Imamura, M. Human macrophage metalloelastase gene expression in colorectal carcinoma and its clinicopathologic significance. *Cancer* **2001**, *91*, 1277–1283, doi:10.1002/1097-0142(20010401)91:7<1277::aid-cncr1129>3.0.co;2-h.
- 94. Shi, H.; Xu, J.M.; Hu, N.Z.; Wang, X.L.; Mei, Q.; Song, Y.L. Transfection of mouse macrophage metalloelastase gene into murine CT-26 colon cancer cells suppresses orthotopic tumor growth, angiogenesis and vascular endothelial growth factor expression. *Cancer Lett.* **2006**, 233, 139–150, doi:10.1016/j.canlet.2005.03.010.
- 95. Xu, Z.; Shi, H.; Li, Q.; Mei, Q.; Bao, J.; Shen, Y.; Xu, J. Mouse macrophage metalloelastase generates angiostatin from plasminogen and suppresses tumor angiogenesis in murine colon cancer. *Oncol. Rep.* **2008**, *20*, 81–88, doi:10.3892/or.20.1.81.

Cancers **2021**, 13, 6226 19 of 21

96. Beurden, P.; Von den Hoff, J. Zymographic techniques for the analysis of matrix metalloproteinases and their inhibitors. *Bio-Techniques* **2005**, *38*, 73–83, doi:10.2144/05381RV01.

- 97. Wu, T.; Li, Y.; Liu, X.; Lu, J.; He, X.; Wang, Q.; Li, J.; Du, X. Identification of high-risk stage II and stage III colorectal cancer by analysis of MMP-21 expression. *J. Surg. Oncol.* **2011**, *104*, 787–791, doi:10.1002/jso.21970.
- 98. Huang, Y.; Li, W.; Chu, D.; Zheng, J.; Ji, G.; Li, M.; Zhang, H.; Wang, W.; Du, J.; Li, J. Overexpression of matrix metalloproteinase-21 is associated with poor overall survival of patients with colorectal cancer. *J. Gastrointest. Surg.* **2011**, *15*, 1188–1194, doi:10.1007/s11605-011-1519-5.
- 99. Zhang, J.; Pan, Q.; Yan, W.; Wang, Y.; He, X.; Zhao, Z. Overexpression of MMP21 and MMP28 is associated with gastric cancer progression and poor prognosis. *Oncol. Lett.* **2018**, *15*, 7776–7782, doi:10.3892/ol.2018.8328.
- 100. Pahwa, S.; Stawikowski, M.J.; Fields, G.B. Monitoring and Inhibiting MT1-MMP during Cancer Initiation and Progression. *Cancers* **2014**, *6*, 416–435, doi:10.3390/cancers6010416.
- 101. Devy, L.; Huang, L.; Naa, L.; Yanamandra, N.; Pieters, H.; Frans, N.; Chang, E.; Tao, Q.; Vanhove, M.; Lejeune, A.; et al. Selective Inhibition of Matrix Metalloproteinase-14 Blocks Tumor Growth, Invasion, and Angiogenesis. *Cancer Res.* **2009**, *69*, 1517–1526, doi:10.1158/0008-5472.CAN-08-3255.
- 102. Duan, F.; Peng, Z.; Yin, J.; Yang, Z.; Shang, J. Expression of MMP-14 and prognosis in digestive system carcinoma: A meta-analysis and databases validation. *J. Cancer* **2020**, *11*, 1141–1150, doi:10.7150/jca.36469.
- 103. Yang, B.; Gao, J.; Rao, Z.; Shen, Q. Clinicopathological and prognostic significance of α5β1-integrin and MMP-14 expressions in colorectal cancer. *Neoplasma* **2013**, *60*, 254–261, doi:10.4149/neo_2013_034.
- 104. Cui, G.; Cai, F.; Ding, Z.; Gao, L. MMP14 predicts a poor prognosis in patients with colorectal cancer. *Hum. Pathol.* **2019**, *83*, 36–42, doi:10.1016/j.humpath.2018.03.030.
- 105. Claesson-Welsh, L. How the matrix metalloproteinase MMP14 contributes to the progression of colorectal cancer. *J. Clin. Investig.* **2020**, 130, 1093–1095, doi:10.1172/JCI135239.
- 106. Decock, J.; Paridaens, R.; Ye, S. Genetic polymorphisms of matrix metalloproteinases in lung, breast and colorectal cancer. *Clin. Genet.* **2008**, *73*, 197–211, doi:10.1111/j.1399-0004.2007.00946.x.
- 107. Langers, A.M.; Verspaget, H.W.; Hommes, D.W.; Sier, C.F. Single-nucleotide polymorphisms of matrix metalloproteinases and their inhibitors in gastrointestinal cancer. *World J. Gastrointest. Oncol.* **2011**, *3*, 79–98, doi:10.4251/wjgo.v3.i6.79.
- 108. Hinoda, Y.; Okayama, N.; Takano, N.; Fujimura, K.; Suehiro, Y.; Hamanaka, Y.; Hazama, S.; Kitamura, Y.; Kamatani, N.; Oka, M. Association of functional polymorphisms of matrix metalloproteinase (MMP)-1 and MMP-3 genes with colorectal cancer. *Int. J. Cancer* 2002, 102, 526–529, doi:10.1002/ijc.10750.
- 109. Kouhkan, F.; Motovali-Bashi, M.; Hojati, Z. The influence of interstitial collagenas-1 genotype polymorphism on colorectal cancer risk in Iranian population. *Cancer Investig.* **2008**, *26*, 836–842, doi:10.1080/07357900801953204.
- 110. Langers, A.M.J.; Sier, C.F.M.; Hawinkels, L.J.A.C.; Kubben, F.J.G.M.; van Duijn, W.; van der Reijden, J.J.; Lamers, C.B.H.W.; Hommes, D.W.; Verspaget, H.W. MMP-2 geno-phenotype is prognostic for colorectal cancer survival, whereas MMP-9 is not. *Br. J. Cancer* 2008, *98*, 1820–1823, doi:10.1038/sj.bjc.6604380.
- 111. Ting, W.-C.; Chen, L.-M.; Pao, J.-B.; Yang, Y.-P.; You, B.-J.; Chang, T.-Y.; Lan, Y.-H.; Lee, H.-Z.; Bao, B.-Y. Genetic Polymorphisms of Matrix Metalloproteinases and Clinical Outcomes in Colorectal Cancer Patients. *Int. J. Med. Sci.* 2013, 10, 1022–1027, doi:10.7150/ijms.6686.
- 112. Dziki, L.; Przybyłowska, K.; Majsterek, I.; Trzciński, R.; Mik, M.; Sygut, A. A/G Polymorphism of the MMP-7 Gene Promoter Region in Colorectal Cancer. *Pol. Przegl. Chir.* **2011**, *83*, 622–626, doi:10.2478/v10035-011-0098-1.
- 113. Fang, W.-L.; Liang, W.; He, H.; Zhu, Y.; Li, S.-L.; Gao, L.-B.; Zhang, L. Association of Matrix Metalloproteinases 1, 7, and 9 Gene Polymorphisms with Genetic Susceptibility to Colorectal Carcinoma in a Han Chinese Population. *DNA Cell Biol.* **2010**, 29, 657–661, doi:10.1089/dna.2010.1017.
- 114. Park, K.S.; Kim, S.J.; Kim, K.H.; Kim, J.C. Clinical characteristics of TIMP2, MMP2, and MMP9 gene polymorphisms in colorectal cancer. *J. Gastroenterol. Hepatol.* **2011**, *26*, 391–397, doi:10.1111/j.1440-1746.2010.06504.x.
- 115. Van Nguyen, S.; Skarstedt, M.; LÖFgren, S.; Zar, N.; Andersson, R.E.; Lindh, M.; Matussek, A.; Dimberg, J.A.N. Gene Polymorphism of Matrix Metalloproteinase-12 and -13 and Association with Colorectal Cancer in Swedish Patients. *Anticancer Res.* **2013**, 33, 3247–3250.
- 116. Tai, J.; Sun, D.; Wang, X.; Kang, Z. Matrix metalloproteinase-8 rs11225395 polymorphism correlates with colorectal cancer risk and survival in a Chinese Han population: A case-control study. *Aging* **2020**, *12*, 19618–19627, doi:10.18632/aging.103930.
- 117. Vandenbroucke, R.E.; Libert, C. Is there new hope for therapeutic matrix metalloproteinase inhibition? *Nat. Rev. Drug Discov.* **2014**, *13*, 904–927, doi:10.1038/nrd4390.
- 118. Raeeszadeh-Sarmazdeh, M.; Do, L.D.; Hritz, B.G. Metalloproteinases and Their Inhibitors: Potential for the Development of New Therapeutics. *Cells* **2020**, *9*, 1313, doi:10.3390/cells9051313.
- 119. Caton, J.G. Evaluation of Periostat for patient management. Compend. Contin. Educ. Dent. 1999, 20, 451, 458-460.
- 120. Bissett, D.; O'Byrne, K.J.; von Pawel, J.; Gatzemeier, U.; Price, A.; Nicolson, M.; Mercier, R.; Mazabel, E.; Penning, C.; Zhang, M.H; et al. Phase III study of matrix metalloproteinase inhibitor prinomastat in non-small-cell lung cancer. *J. Clin. Oncol.* 2005, 23, 842–849, doi:10.1200/JCO.2005.03.170.
- 121. Scatena, R. Prinomastat, a hydroxamate-based matrix metalloproteinase inhibitor. A novel pharmacological approach for tissue remodelling-related diseases. *Expert Opin. Investig. Drugs* **2000**, *9*, 2159–2165, doi:10.1517/13543784.9.9.2159.

Cancers **2021**, 13, 6226 20 of 21

122. Hande, K.R.; Collier, M.; Paradiso, L.; Stuart-Smith, J.; Dixon, M.; Clendeninn, N.; Yeun, G.; Alberti, D.; Binger, K.; Wilding, G. Phase I and pharmacokinetic study of prinomastat, a matrix metalloprotease inhibitor. *Clin. Cancer Res.* **2004**, *10*, 909–915, doi:10.1158/1078-0432.ccr-0981-3.

- 123. Yang, J.-S.; Lin, C.-W.; Su, S.-C.; Yang, S.-F. Pharmacodynamic considerations in the use of matrix metalloproteinase inhibitors in cancer treatment. *Expert Opin. Drug Metab. Toxicol.* **2015**, *12*, 191–200, doi:10.1517/17425255.2016.1131820.
- 124. Shah, M.A.; Starodub, A.; Sharma, S.; Berlin, J.; Patel, M.; Wainberg, Z.A.; Chaves, J.; Gordon, M.; Windsor, K.; Brachmann, C.B; et al. Andecaliximab/GS-5745 Alone and Combined with mFOLFOX6 in Advanced Gastric and Gastroesophageal Junction Adenocarcinoma: Results from a Phase I Study. *Clin. Cancer Res.* 2018, 24, 3829–3837, doi:10.1158/1078-0432.CCR-17-2469.
- 125. Sandborn, W.J.; Bhandari, B.R.; Randall, C.; Younes, Z.H.; Romanczyk, T.; Xin, Y.; Wendt, E.; Chai, H.; McKevitt, M.; Zhao, S.; et al. Andecaliximab [Anti-matrix Metalloproteinase-9] Induction Therapy for Ulcerative Colitis: A Randomised, Double-Blind, Placebo-Controlled, Phase 2/3 Study in Patients With Moderate to Severe Disease. *J. Crohn's Colitis* 2018, 12, 1021–1029, doi:10.1093/ecco-jcc/jjy049.
- 126. Murphy, G. Tissue inhibitors of metalloproteinases. Genome Biol. 2011, 12, 233-233, doi:10.1186/gb-2011-12-11-233.
- 127. Li, K.; Tay, F.R.; Yiu, C.K.Y. The past, present and future perspectives of matrix metalloproteinase inhibitors. *Pharmacol. Ther.* **2020**, 207, 107465, doi:10.1016/j.pharmthera.2019.107465.
- 128. Melendez-Zajgla, J.; Del Pozo, L.; Ceballos, G.; Maldonado, V. Tissue Inhibitor of Metalloproteinases-4. The road less traveled. *Mol. Cancer* 2008, 7, 85, doi:10.1186/1476-4598-7-85.
- 129. Hayden, D.M.; Forsyth, C.; Keshavarzian, A. The role of matrix metalloproteinases in intestinal epithelial wound healing during normal and inflammatory states. *J. Surg. Res.* **2011**, *168*, 315–324, doi:10.1016/j.jss.2010.03.002.
- 130. Song, G.; Xu, S.; Zhang, H.; Wang, Y.; Xiao, C.; Jiang, T.; Wu, L.; Zhang, T.; Sun, X.; Zhong, L.; et al. TIMP1 is a prognostic marker for the progression and metastasis of colon cancer through FAK-PI3K/AKT and MAPK pathway. *J. Exp. Clin. Cancer Res.* 2016, 35, 148, doi:10.1186/s13046-016-0427-7.
- 131. Noël, A.; Jost, M.; Maquoi, E. Matrix metalloproteinases at cancer tumor-host interface. Semin. Cell Dev. Biol. 2008, 19, 52–60, doi:10.1016/j.semcdb.2007.05.011.
- 132. Lu, X.; Duan, L.; Xie, H.; Lu, X.; Lu, D.; Lu, D.; Jiang, N.; Chen, Y. Evaluation of MMP-9 and MMP-2 and their suppressor TIMP-1 and TIMP-2 in adenocarcinoma of esophagogastric junction. *OncoTargets Ther.* **2016**, *9*, 4343–4349, doi:10.2147/OTT.S99580.
- 133. Groblewska, M.; Mroczko, B.; Gryko, M.; Pryczynicz, A.; Guzińska-Ustymowicz, K.; Kędra, B.; Kemona, A.; Szmitkowski, M. Serum levels and tissue expression of matrix metalloproteinase 2 (MMP-2) and tissue inhibitor of metalloproteinases 2 (TIMP-2) in colorectal cancer patients. *Tumor Biol.* **2014**, *35*, 3793–3802, doi:10.1007/s13277-013-1502-8.
- 134. Wang, W.; Li, D.; Xiang, L.; Lv, M.; Tao, L.; Ni, T.; Deng, J.; Gu, X.; Masatara, S.; Liu, Y.; et al. TIMP-2 inhibits metastasis and predicts prognosis of colorectal cancer via regulating MMP-9. *Cell Adhes. Migr.* **2019**, 13, 273–284, doi:10.1080/19336918.2019.1639303.
- 135. Huang, H.-L.; Liu, Y.-M.; Sung, T.-Y.; Huang, T.-C.; Cheng, Y.-W.; Liou, J.-P.; Pan, S.-L. TIMP3 expression associates with prognosis in colorectal cancer and its novel arylsulfonamide inducer, MPT0B390, inhibits tumor growth, metastasis and angiogenesis. *Theranostics* **2019**, *9*, 6676–6689, doi:10.7150/thno.34020.
- 136. Lin, H.; Zhang, Y.; Wang, H.; Xu, D.; Meng, X.; Shao, Y.; Lin, C.; Ye, Y.; Qian, H.; Wang, S. Tissue inhibitor of metalloprotein-ases-3 transfer suppresses malignant behaviors of colorectal cancer cells. *Cancer Gene Ther.* **2012**, *19*, 845–851, doi:10.1038/cgt.2012.70.
- 137. Soheilifar, M.H.; Grusch, M.; Keshmiri Neghab, H.; Amini, R.; Maadi, H.; Saidijam, M.; Wang, Z. Angioregulatory microRNAs in Colorectal Cancer. *Cancers* **2019**, *12*, 381–387, doi:10.3390/cancers12010071.
- 138. Wu, J.; Wu, G.; Lv, L.; Ren, Y.; Zhang, X.; Xue, Y.; Li, G.; Lu, X.; Sun, Z.; Tang, K. MicroRNA-34a inhibits migration and invasion of colon cancer cells via targeting to Fra-1. *Carcinogenesis* **2012**, *33*, 519–528, doi:10.1093/carcin/bgr304.
- 139. Abba, M.; Patil, N.; Allgayer, H. MicroRNAs in the Regulation of MMPs and Metastasis. *Cancers* **2014**, *6*, 625–645, doi:10.3390/cancers6020625.
- 140. Shen, K.; Liang, Q.; Xu, K.; Cui, D.; Jiang, L.; Yin, P.; Lu, Y.; Li, Q.; Liu, J. MiR-139 inhibits invasion and metastasis of colorectal cancer by targeting the type I insulin-like growth factor receptor. *Biochem. Pharmacol.* **2012**, *84*, 320–330, doi:10.1016/j.bcp.2012.04.017.
- 141. Tang, W.; Zhu, Y.; Gao, J.; Fu, J.; Liu, C.; Liu, Y.; Song, C.; Zhu, S.; Leng, Y.; Wang, G.; et al. MicroRNA-29a promotes colorectal cancer metastasis by regulating matrix metalloproteinase 2 and E-cadherin via KLF4. *Br. J. Cancer* **2014**, *110*, 450–458, doi:10.1038/bjc.2013.724.
- 142. Leng, Y.; Chen, Z.; Ding, H.; Zhao, X.; Qin, L.; Pan, Y. Overexpression of microRNA-29b inhibits epithelial-mesenchymal transition and angiogenesis of colorectal cancer through the ETV4/ERK/EGFR axis. *Cancer Cell Int.* **2021**, 21, 17, doi:10.1186/s12935-020-01700-2.
- 143. Yu, B.; Liu, X.; Chang, H. MicroRNA-143 inhibits colorectal cancer cell proliferation by targeting MMP7. *Minerva Med.* **2017**, 108, 13–19, doi:10.23736/S0026-4806.16.04651-6.
- 144. Schwarzmueller, L.; Bril, O.; Vermeulen, L.; Léveillé, N. Emerging Role and Therapeutic Potential of lncRNAs in Colorectal Cancer. *Cancers* **2020**, *12*, 3843, doi:10.3390/cancers12123843.
- 145. Tian, L.; Zhao, Z.F.; Xie, L.; Zhu, J.P. Taurine up-regulated 1 accelerates tumorigenesis of colon cancer by regulating miR-26a-5p/MMP14/p38 MAPK/Hsp27 axis in vitro and in vivo. *Life Sci.* **2019**, 239, 117035, doi:10.1016/j.lfs.2019.117035.

Cancers 2021, 13, 6226 21 of 21

146. Sun, N.; Zhang, G.; Liu, Y. Long non-coding RNA XIST sponges miR-34a to promotes colon cancer progression via Wnt/β-catenin signaling pathway. *Gene* **2018**, *665*, 141–148, doi:10.1016/j.gene.2018.04.014.

- 147. Lv, H.; Zhou, D.; Liu, G. LncRNA LINC00963 promotes colorectal cancer cell proliferation and metastasis by regulating miR--1281 and TRIM65. *Mol. Med. Rep.* **2021**, 24, 781, doi:10.3892/mmr.2021.12421.
- 148. Duan, Y.; Fang, Z.; Shi, Z.; Zhang, L. Knockdown of lncRNA CCEPR suppresses colorectal cancer progression. *Exp. Ther. Med.* **2019**, *18*, 3534–3542, doi:10.3892/etm.2019.7942.
- 149. Pan, Y.; Zhu, L.; Pu, J.; Wang, W.; Qian, W. lncRNA PCA3 plays a key role in colon cancer occurrence and development. *Arch. Med Sci.* **2020**, doi:10.5114/aoms.2020.100674.